

For Research Use Only. NOT for Use in Diagnostic Procedures.

Extraction of NBOMe Designer Drugs from Oral Fluid Using ISOLUTE® SLE+ Prior to Analysis by LDTD-MS/MS

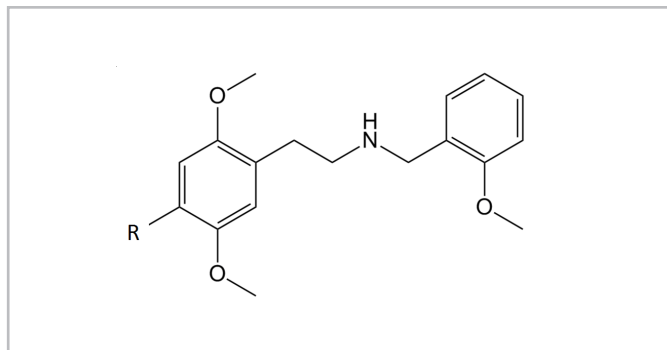
Megan R. Record^{1,2}, Serge Auger³, Pierre Picard³, Frank Kero⁴, Victor Vandell⁴, Alex Birsan³, Amanda L.A. Mohr², Karen S. Scott^{1,2}

1. Arcadia University, 450 S. Easton Road, Glenside, PA 19038

2. The Center for Forensic Science Research and Education, 2300 Stratford Avenue, Willow Grove, PA 190902, USA

3. Phytrolix Technologies, Québec, Canada

4. Biotage, 10430 Harris Oaks Blvd., Suite C, Charlotte North Carolina 28269, USA



Substitution	Analyte
R = Br	25B-NBOMe
R = Cl	25C-NBOMe
R = CH ₃	25D-NBOMe
R = C ₂ H ₅	25E-NBOMe
R = H	25H-NBOMe
R = I	25I-NBOMe
R = N ₂ O	25N-NBOMe
R = SC ₂ H ₅	25T2-NBOMe

Figure 1. Structures of the NBOMes studied

Introduction

This application note describes the extraction of a suite of NBOMe designer drugs from oral fluid matrix with analysis by laser diode thermal desorption (LDTD)-MS/MS.

NBOMes (**Figure 1**) are a class of novel psychoactive substances (NPSs) marketed as “legal highs.” NBOMes are the N-Benzyl-Oxy-Methyl derivatives of previously known phenethylamines in the 2C series. The 2C series contain methoxy groups on the 2 and 5 positions of a benzene ring of the phenethylamine backbone structure. NBOMes also contain a 2-methoxybenzyl on the nitrogen backbone, which results in increased substitution and ultimately potency. Oral fluid contains primarily the parent compound, which is most commonly associated with the pharmacological effects, making it an ideal matrix for the detection of NBOMe analytes.

ISOLUTE® SLE+ Supported Liquid Extraction plates and columns offer an efficient alternative to traditional liquid-liquid extraction (LLE) for bioanalytical sample preparation, providing high analyte recoveries, no emulsion formation, and significantly reduced sample preparation.

Analytes

25B-NBOMe, 25C-NBOMe, 25D-NBOMe, 25E-NBOMe, 25H-NBOMe, 25I-NBOMe, 25N-NBOMe, 25T2-NBOMe

Sample Preparation Procedure

Optimized methodology for extending the dynamic range of the analysis is provided below.

Concentration Range 1: 25–1000 ng/mL

Sample Pre-treatment: To 150 µL of sample, add 25I-NBOMe-D3 internal standard (15 µL, (1000 ng /mL in MeOH), and ammonium hydroxide (NH₄OH, 0.1%, 10 µL). Mix.

Format: ISOLUTE® SLE+ 400 µL Sample Volume columns, part number 820-0055-B

Sample loading: Load pre-treated oral fluid (175 µL, as above) onto the column and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.

Analyte extraction: Apply methyl-tert-butyl-ether (MTBE, 4 x 500 µL) and allow to flow under gravity for 5 minutes. Apply vacuum or positive pressure (5–10 seconds) to complete elution.

LAZWell Spotting: Deposit 3 µL of elution phase onto the LazWell plate and let dry.

Concentration Range 2: 0.5–100 ng/mL

Sample Pre-treatment:	To 300 µL of sample, add 25I-NBOMe-D3 internal standard (30 µL, (100 ng /mL in MeOH), and ammonium hydroxide (NH ₄ OH, 0.1%, 20 µL). Mix.
Format:	ISOLUTE® SLE+ 400 µL Sample Volume columns, part number 820-0055-B
Sample loading:	Load pre-treated oral fluid (350 µL, as above) onto the column and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.
Analyte extraction:	Apply MTBE (4 x 500 µL) and allow to flow under gravity for 5 minutes. Apply vacuum or positive pressure (5–10 seconds) to complete elution.
LAZWell Spotting:	Deposit 6 µL of elution phase onto the LazWell plate and let dry.

Mass Spectrometry Conditions

Following sample preparation, a small volume is transferred and dried into a well cavity (see LAZWell Spotting, above). The analytes of interest were vaporized indirectly by thermal action and ionized by APCI. The time required was less than **9 seconds per sample**.

The LDTD-MS was coupled to a SCIEX 5500 QTRAP (**Figure 4**). The MS/MS instrumentation was operated in multi-reaction monitoring mode (MRM). The optimized compound selective parameters are provided in **Table 1**.

Table 1. Compound selective MS/MS parameters

MRM	Q1 m/z	Q3 m/z	Dwell ms	Compound	Collision Energy eV
1	302.2	120.6	5	25H-121	22
2	302.2	165.1	5	25H-165	22
3	316.0	120.6	5	25D-121	22
4	316.0	179.0	5	25D-179	22
5	330.1	120.7	5	25E-120	22
6	330.1	192.9	5	25E-192	22
7	336.1	120.7	5	25C-121	22
8	336.1	90.9	5	25C-91	22
9	347.2	120.6	5	25N-121	22
10	347.2	90.8	5	25N-91	22
11	348.2	120.7	5	25T2-121	22
12	348.2	211.1	5	25T2-211	22
13	380.2	120.8	5	25B-121	22
14	380.2	91.0	5	25B-91	22
15	428.1	120.7	5	25I-121	22
16	428.1	272.1	5	25I-272	22
17	431.1	124.0	5	25I-D3-124-IS	22

Results

Linearity was determined for two of the analytes of interest (R=Br, I) to verify method performance. The results are reported in **Figures 2 and 3**.

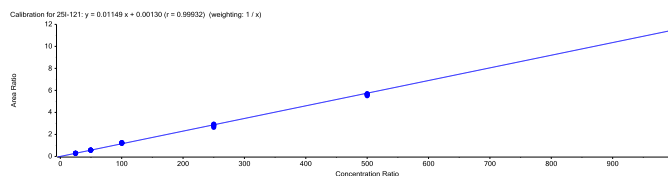


Figure 2. Calibration curve for 25B-NBOMe (calibration range 1: 25-1000 ng/mL)

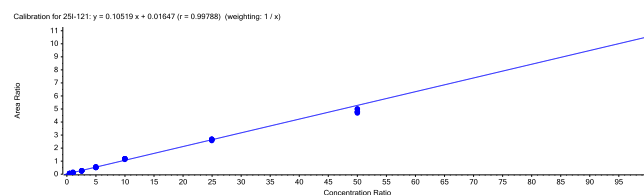


Figure 3. Calibration curve for 25I-NBOMe (calibration range 2: 0.5-100 ng/mL)

A set of oral fluid specimens was prepared at the Center for Forensic Science Research and Education. The samples were collected with Salivettes and fortified to different concentration levels. The samples were submitted blind for analysis. Results are shown in **Figure 4**.

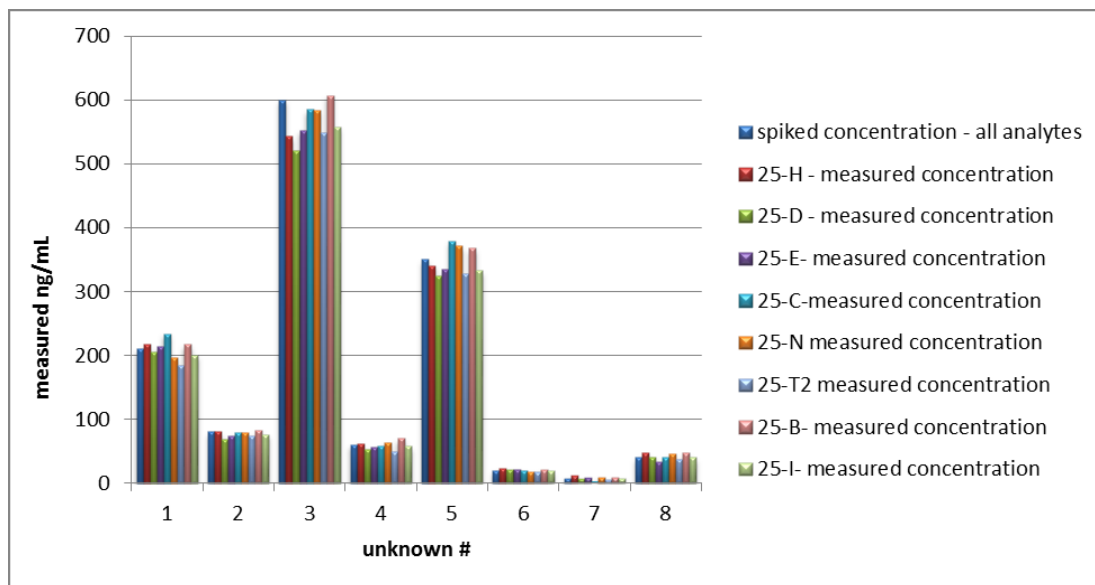


Figure 4. Unknown sample set fortified to 8 concentration levels for a panel of NBOMe designer drugs, and extracted using the method detailed in this application note.

Conclusions

In this application note, SLE-LDTD-MS is demonstrated to be a viable workflow solution for the screening of NBOMe designer drugs in oral fluids. ISOLUTE® SLE+ columns proved effective in minimizing matrix effects by producing clean extracts and subsequent LDTD-MS/MS allowed ultrafast high throughput analysis at 9 seconds per sample.

The method demonstrated good sensitivity with LLOQ values determined at 0.5 ng/mL for 25B-NBOMe and 25I-NBOMe. This method demonstrated acceptable precision and accuracy across the calibration range of interest, with linearity of $r^2 > 0.99$ for both analytes and reproducibility with CV < 10% (n=4) (<15% at LLOQ).

This application note is based on the poster 'A Novel SLE-LDTD-MS/MS Method for the Screening of NBOMe Designer Drugs in Oral Fluid' presented at TAMS 2014, Research Triangle Park, NC, USA and NEAFS 2014, Hershey, PA, USA.

Ordering Information

Part Number	Description	Quantity
820-0055-B	ISOLUTE® SLE+ 400 µL Supported Liquid Extraction Columns	50
PPM-48	Biotage® Positive Pressure Manifold 48 Position	1

EUROPE

Main Office: +46 18 565900
Toll Free: +800 18 565710
Fax: +46 18 591922
Order Tel: +46 18 565710
Order Fax: +46 18 565705
order@biotage.com
Support Tel: +46 18 56 59 11
Support Fax: +46 18 56 57 11
eu-1-pointsupport@biotage.com

NORTH & LATIN AMERICA

Main Office: +1 704 654 4900
Toll Free: +1 800 446 4752
Fax: +1 704 654 4917
Order Tel: +1 704 654 4900
Order Fax: +1 434 296 8217
ordermailbox@biotage.com
Support Tel: +1 800 446 4752
Outside US: +1 704 654 4900
us-1-pointsupport@biotage.com

JAPAN

Tel: +81 3 5627 3123
Fax: +81 3 5627 3121
jp_order@biotage.com
jp-1-pointsupport@biotage.com

CHINA

Tel: +86 21 2898 6655
Fax: +86 21 2898 6153
cn_order@biotage.com
cn-1-pointsupport@biotage.com

To locate a distributor, please visit our website at www.biotage.com

Part Number: AN834.V.1

© 2014 Biotage. All rights reserved. No material may be reproduced or published without the written permission of Biotage. Information in this document is subject to change without notice and does not represent any commitment from Biotage. E&OE. Product and company names mentioned herein may be trademarks or registered trademarks and/or service marks of their respective owners, and are used only for explanation and to the owners' benefit, without intent to infringe. **FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.**

For more information visit www.biotage.com.