

Strategies for Extraction of Drugs from Whole Blood

Stephanie J. Marin, Mario Merida, Jillian Neifeld, Jeremy Smith, Mohamed Youssef, Elena Gairloch
Biotage, 10430 Harris Oaks Blvd., Suite C, Charlotte North Carolina 28269, USA



Introduction

Whole blood is often the specimen of choice for detection of drugs in the forensic laboratory. Samples are typically extracted to remove interferences and isolate the analytes of interest prior to GC-MS or LC-MS/MS analysis. Sample clean-up reduces interferences that can suppress MS response and decrease column lifetime or clog chromatography instrumentation. Here two “pass through” solid phase extraction techniques (protein removal/phospholipid depletion and dual mode extraction) and supported liquid extraction were evaluated for a group of about 100 compounds in whole blood. These DOA compounds included drugs from multiple drug classes: anticonvulsants, barbiturates, benzodiazepines, opioids, antidepressants, stimulants, hallucinogens, cannabinoids and antipsychotics.

Methods

All standards were purchased from Ceriliant (Round Rock, TX). HPLC grade water, methanol (MeOH), dichloromethane (DCM), formic acid (FA), ammonium hydroxide (NH₄OH) methyl tert-butyl ether (MTBE), isopropanol (IPA), ethyl acetate (EtOAc) and acetonitrile (ACN) were purchased from Sigma Aldrich (St. Louis, MO). Biotage® ISOLUTE® HYDRO DME+ dual mode extraction plates (970-0400-PZ01), ISOLUTE® PLD+ protein removal/phospholipid depletion extraction plates (918-0050-P01) and ISOLUTE® SLE+ 400 µL supported liquid extraction plates (820-0400-P01) were used for the extractions. Samples were processed using a Biotage® PRESSURE+ 96 position positive pressure manifold (PPM-96), and a Biotage® SPE Dry 96 (SD-9300-DHS-NA). LC columns were generously provided by Restek.

Whole blood was provided by UTAK (Valencia, CA). Fortified samples were prepared at 100 ng/mL for all compounds. An unextracted no matrix standard prepared in reconstitution solvent was also prepared at the same concentration and analyzed to calculate process efficiency (1). After extraction, samples were analyzed by LC-MS/MS using a Nexera UPLC (Shimadzu, Kyoto Prefecture, Japan) and a SCIEX 5500 triple quadrupole mass spectrometer with Turbo Ionspray® Ion Interface (Sciex, Framingham, MA) in scheduled MRM mode.

Table 1. LC Parameters

Column	Restek Raptor Biphenyl 2.7 µm, 50 x 3.0 mm		
MPA	0.1% Formic Acid (aq)		
MPB	0.1% Formic Acid in MeOH		
Flow Rate	0.45 mL/min		
Column Temp	40 °C		
Sample Temp	20 °C		
Injection Volume	2.5 µL		

The LC gradient started with 95% MPA and gradually decreased to 5% MPA over a 9.0 minute total run time.

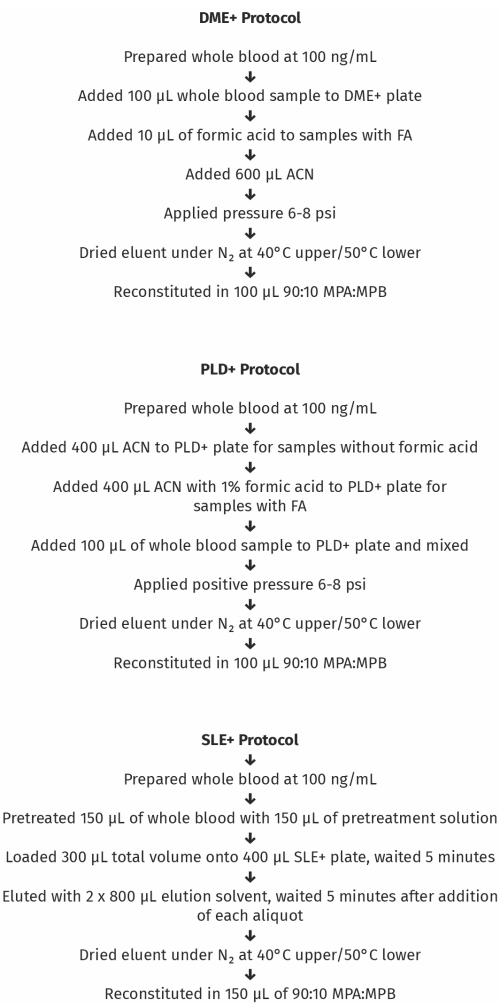
Optimized source parameters are shown in Table 2 (sMRM transition parameters not shown, but available upon request). Retention window for sMRM was set at 45 seconds with a target scan time at 2.85 seconds.

Table 2. MS Parameters

Ionization Spray Voltage	+1500(V)	CAD	Medium
Source Temp	600 °C	GS1	50
Curtain	30 (V)	GS2	70

Extraction Protocols

Samples were extracted following the manufacturer’s protocol. ISOLUTE® SLE+ samples were pretreated with 1% formic acid, water, or 0.5M NH₄OH prior to extraction. Samples were extracted and analyzed in triplicate and average values are reported. A negative control and an extraction blank were also extracted and analyzed.



Results

Results for selected drug classes are in Figures 1 and 2.

Figure 1. Process Efficiency for PLD+ and DME+

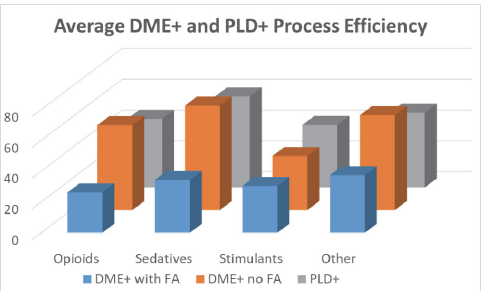
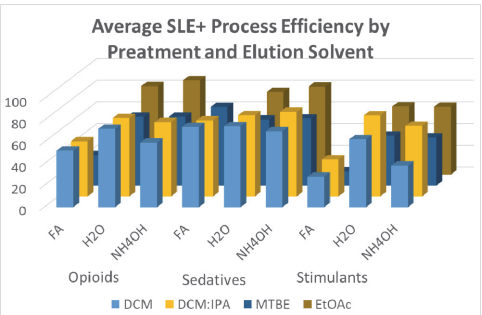


Figure 2. Process Efficiency SLE+



Opioids included 6-AM, buprenorphine, codeine, EDDP, fentanyl, hydrocodone, hydromorphone, meperidine, methadone, morphine, naloxone, norbuprenorphine, norfentanyl, oxycodone, oxymorphone, tapentadol, tramadol. Sedatives consisted of 7-aminoclozapem, alprazolam, clonazepam, diazepam, lorazepam, midazolam, nordiazepam, oxazepam, phenobarbital, secobarbital, temazepam, and triazolam. Stimulants were comprised of amphetamine, benzoyllecgonine, cocaethylene, cocaine, mCPP, MDA, MDEA, MDMA, methamphetamine, methcathinone, and ritalinic acid. “Other” was a selection from the remaining drug classes: amitriptyline, aripiprazole, atomoxetine, bupropion, buspirone, carbamazepine, carisoprodol, chlorpheniramine, chlorpromazine, clomipramine, clonidine, clozapine, cyclobenzaprine, dextromethorphan, duloxetine, fluoxetine, haloperidol, imipramine, ketamine, lamotrigine, levetiracetam, lidocaine, meprobamate, methaqualone, mirtazapine, nortriptyline, olanzapine, oxcarbazepine, paroxetine, phenytoin, quetiapine, risperidone, sertraline, topiramate, trazodone, trimipramine and venlafaxine.

The whole blood used in this study contained high amounts of gabapentin, so process efficiency could not be determined.

Results (continued)

Table 3. Process Efficiency Data for DME+ and PLD+

Value	Opioids			Sedatives			Stimulants			Other		
	DME+ DME+ FA PLD+	DME+ DME+ FA PLD+	DME+ DME+ FA PLD+	DME+ DME+ FA PLD+	DME+ DME+ FA PLD+	DME+ DME+ FA PLD+	DME+ DME+ FA PLD+	DME+ DME+ FA PLD+	DME+ DME+ FA PLD+	DME+ DME+ FA PLD+	DME+ DME+ FA PLD+	DME+ DME+ FA PLD+
min	31.8	2.1	16.7	13.8	0.7	30.3	0.1	1.8	20.9	20.9	0.1	14.2
max	99.6	89.1	87.1	129.7	129.7	129.7	81.3	62.8	74.3	87.7	69.5	74.9
average	55.2	26.2	44.6	67.9	34.4	51.1	35.2	30.4	40.6	61.7	37.4	48.6
median	53.9	24.6	41.2	62.3	34.9	52.6	32.0	27.9	38.1	62.4	37.6	50.3

Table 4. Process Efficiency Data for SLE+

Pre-treatment	Value	Opioids				Sedatives			
		DCM	DCM:IPA	MTBE	EtOAc	DCM	DCM:IPA	MTBE	EtOAc
FA	min	3.2	2.5	1.7	-	27.9	26.9	39.5	-
	max	94.5	101.1	92.5	-	92.1	94.1	91.2	-
	average	52.1	50.7	27.9	-	74.0	70.0	72.3	-
	median	65.9	53.3	13.7	-	76.3	75.2	73.9	-
H2O	min	23.0	17.4	17.4	59.0	62.4	61.1	50.3	64.3
	max	100.7	121.3	82.6	99.6	95.3	92.2	73.2	90.2
	average	72.2	72.1	63.3	81.5	74.6	74.8	60.8	76.1
	median	77.9	76.3	63.5	83.5	74.6	74.3	61.8	78.6
NH ₄ OH	min	20.2	33.2	17.4	44.8	7.7	59.6	50.7	66.6
	max	91.3	127.5	82.6	120.9	88.4	92.0	71.1	91.9
	average	59.6	68.4	63.3	87.0	70.0	77.8	61.6	81.1
	median	65.4	68.7	63.5	86.7	72.5	80.4	63.8	84.6
Pre-treatment	Value	Stimulants				Other			
		DCM	DCM:IPA	MTBE	EtOAc	DCM	DCM:IPA	MTBE	EtOAc
FA	min	0.0	2.8	0.0	-	41.2	26.9	3.4	-
	max	94.6	99.3	60.4	-	100.5	115.3	103.7	-
	average	28.2	34.3	12.6	-	75.0	81.5	56.1	-
	median	14.2	19.3	4.4	-	80.0	82.0	63.3	-
H2O	min	0.0	4.8	0.0	0.4	20.4	31.5	2.4	29.2
	max	87.2	107.2	73.2	89.3	114.9	120.4	89.7	106.4
	average	62.6	74.6	45.9	62.7	78.7	76.6	65.0	79.4
	median	72.6	86.4	56.1	76.2	82.2	78.7	67.4	80.6
NH ₄ OH	min	0.0	3.6	0.0	0.3	12.1	23.0	2.7	24.6
	max	70.1	105.5	66.6	89.0	98.2	127.6	93.0	102.6
	average	38.4	65.0	44.2	62.4	69.9	75.2	66.6	81.0
	median	51.3	73.2	54.8	78.5	71.8	78.5	69.9	84.0

Samples pretreated with formic acid and extracted with PLD+ were dirty and not injected. Samples pretreated with formic acid and extracted using SLE+ and ethyl acetate were dirty and not injected.

Conclusions

- » Acetonitrile with formic acid can’t be used as an extraction solvent for whole blood on ISOLUTE® PLD+, but whole blood can be treated with formic acid prior to extraction using ISOLUTE® HYDRO DME+
- » Ritalinic acid and pregabalin showed low recovery under all conditions, but were detected
- » Best results for basic analytes were without formic acid on ISOLUTE® HYDRO DME+ and using NH₄OH pretreatment using ISOLUTE® SLE+
- » Acidic drugs had the highest process efficiencies using formic acid pretreatment on ISOLUTE® HYDRO DME+ and ISOLUTE® SLE+
- » Overall the best process efficiencies for most compounds used neutral or basic pretreatment and ISOLUTE® SLE+
- » Minimal cleanup of whole blood with pass through SPE or SLE+ resulted in very clean samples and detection of most compounds

References

1. Matuszewski BK et al, *Anal. Chem.* 75, 3019-3030 (2003)