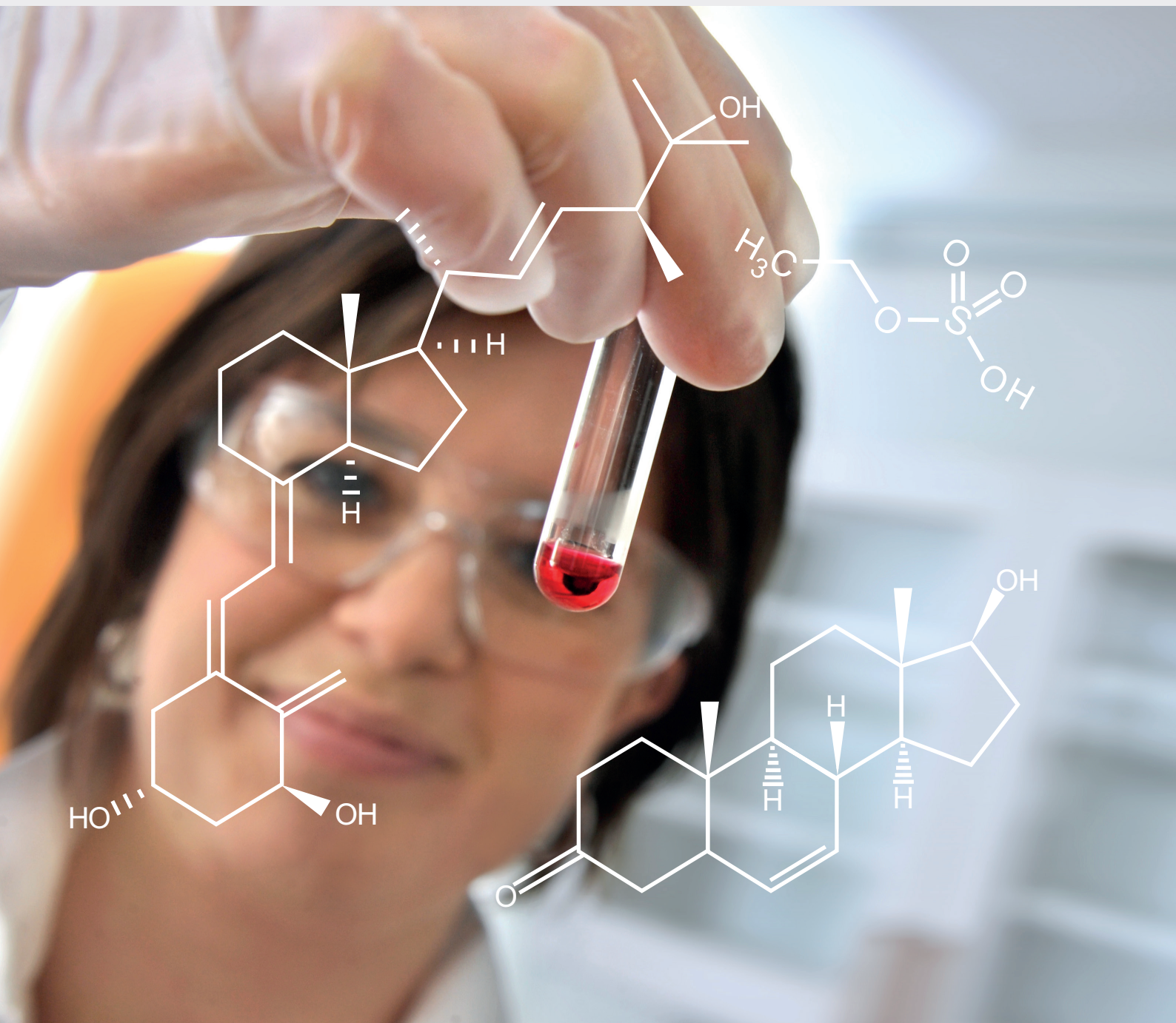


Clinical Applications

Compendium



For Research Use Only. NOT for Use in Diagnostic Procedures.

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Introduction

Welcome to the Biotage Clinical Applications Compendium

This compendium highlights a selection of clinical sample preparation applications from Biotage. Whether you need targeted methods for analytes such as Vitamin D metabolites in serum, or methods suitable for extraction of a wide panel of drugs and metabolites from urine, sample preparation before analysis is essential.

Sample preparation to remove matrix components such as proteins, phospholipids and salts from your biological fluid samples can improve analyte sensitivity, method robustness and improve the quality of analytical data, as well as reducing instrument downtime.



Automated Using Biotage® Extrahera™

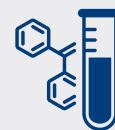
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Choosing the 'right' sample preparation approach for every assay depends on a multitude of factors, including analyte(s), matrix type, analytical methodology, speed, cost per sample, extract cleanliness and sensitivity requirements and throughput needs. The methods outlined in this compendium use the range of Biotage sample preparation products:

- » EVOLUTE® EXPRESS Solid Phase Extraction products
- » ISOLUTE® SLE+ Supported Liquid Extraction products
- » ISOLUTE® PLD+ Protein and Phospholipid Removal products
- » ISOLUTE® PPT+ Protein Precipitation products

Detailed extraction and analytical methodology can be found in the full application notes, downloadable from www.biotage.com.



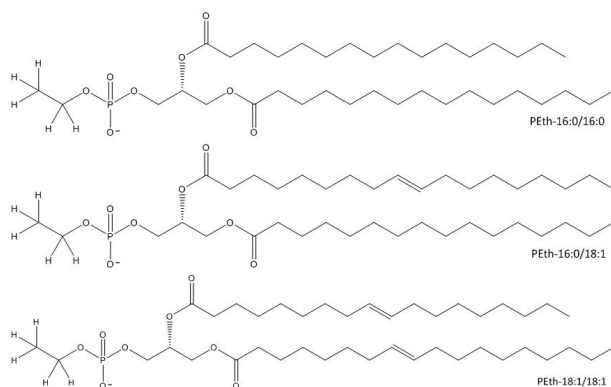


Alcohol Biomarkers

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AN755	Simultaneous Extraction of Ethyl Glucuronide and Ethyl Sulfate from Urine with EVOLUTE® EXPRESS AX Prior to LC-MS/MS analysis	page 3

AN876

Extraction of Phosphatidylethanol (PEth) Species from Whole Blood Using ISOLUTE® SLE+ Prior to HPLC-MS/MS Analysis



Chemical structures of three common PEth species.

Analytes

1,2-dipalmitoyl-sn-glycero-3-phosphoethanol (PEth-16:0/16:0), 1-palmitoyl-2-oleoyl-sn-glycero-3-phosphoethanol (PEth-16:0/18:1), 1,2-dioleoyl-sn-glycero-3-phosphoethanol (PEth-18:1/18:1).

Format

ISOLUTE® SLE+ 200 µL Supported Liquid Extraction plate, part number 820-0200-P01.

Matrices

Whole blood.

Sample Preparation Method

To whole blood (20 µL), add 6.25% (v/v) aqueous ammonium hydroxide in 30% aqueous methanol (300 µL). Add internal standard. Mix thoroughly and allow to equilibrate. Load 140 µL of the pre-treated sample (equivalent to 8.75 µL whole blood). Apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.

Elute analytes with ethyl acetate (750 µL).

Post Extraction

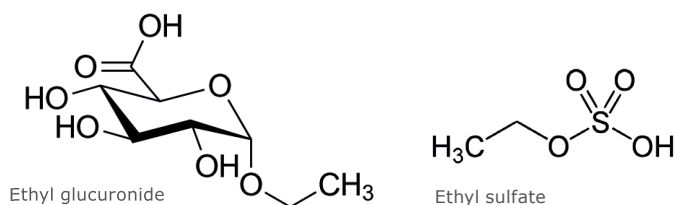
Dry the extract in a stream of air or nitrogen at 40 °C. Reconstitute the extracts with 150 µL of mobile phase and mix thoroughly before analysis.

Summary of Results

Recoveries range from 85–89%, with RSDs ~5%. LOQ <30 nmol/L.

AN818

A No-Drydown SPE Method for Biomarkers of Alcohol Consumption in Human Urine Using ISOLUTE® NH2 SPE Columns Prior to LC-MS/MS



Structures of ethyl glucuronide and ethyl sulfate.

Analytes

EtG (Ethyl glucuronide) and EtS (Ethyl sulfate).

Format

ISOLUTE® NH2 100 mg/1 mL columns, part number 470-0010-A.



Matrices

Urine

Sample Preparation Method

Dilute urine sample (100 µL) with acetonitrile (1 mL) and add 6M HCl (50 µL). Add internal standard as required and mix. Condition columns with methanol (2 mL), then equilibrate with water (2 mL) followed by acetonitrile containing 0.2% (v/v) acetic acid (2 mL). Load entire sample at a flow rate of 1 mL/min. Dry the column and elute interferences with hexane (1 mL). Dry columns for 10 mins under positive pressure. Elute analytes with 10 mM ammonium formate/formic acid (pH 3, 2 x 750 µL).

Post Extraction

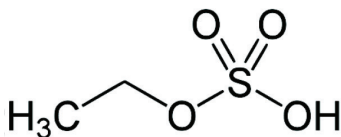
The extract can be injected directly into the analytical system without an additional evaporation step.

Summary of Results

This ISOLUTE NH2 method was able to differentiate between patient positive and patient negative samples over a range of clinical interest.

AN755

Simultaneous Extraction of Ethyl Glucuronide and Ethyl Sulfate from Urine with EVOLUTE® EXPRESS AX Prior to LC-MS/MS Analysis



Structure of Ethyl Sulfate.

Analytes

EtG (Ethyl glucuronide) and EtS (Ethyl sulfate).

Format

EVOLUTE® EXPRESS AX 100 mg/3 mL columns, part number 613-0010-BXG.



Matrices

Urine

Sample Preparation Method

Dilute urine samples with acetonitrile (1:9, v/v) and add internal standard as required. Mix. Condition columns with methanol (3 mL) and equilibrate with water (3 mL) followed by acetonitrile (3 mL). Load the pre-treated sample (2 mL). Elute interferences with acetonitrile (3 mL) followed by methanol (3 mL). Elute analytes with 2% HCl in acetonitrile (3 mL).

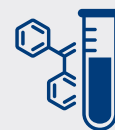
Post Extraction






Evaporate to dryness at 40 °C and reconstitute with HPLC grade water (250 µL). Vortex mix and add acetonitrile (250 µL), vortex mix again before analysis.

Summary of Results

Analyte recoveries of >90% with LOQ of 10 ng/mL for EtG and 2 ng/mL for EtS.

Biomarkers



AN861	Extraction of Methylmalonic Acid from Serum Using ISOLUTE® PPT+ Protein Precipitation Plates Prior to LC-MS/MS Analysis		page 5
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AN737	Extraction of Organophosphate Pesticide Metabolites From Urine Using EVOLUTE® EXPRESS WAX		page 8

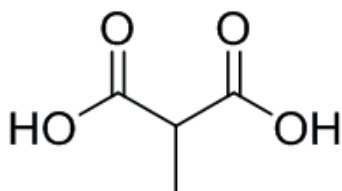


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AN861

Extraction of Methylmalonic Acid from Serum Using ISOLUTE® PPT+ Protein Precipitation Plates Prior to LC-MS/MS Analysis



Structure of methylmalonic acid (MMA).

Analytes

Methylmalonic acid (MMA).

Format

ISOLUTE® PPT+ Protein Precipitation plate, part number 120-2040-P01.

Matrices

Serum.

Sample Preparation Method

To serum (100 µL), add 10 µL of ISTD (10 ng/µL). Mix and allow to equilibrate. Place extraction plate in manifold, with an appropriate collection plate in position. Add 800 µL of acetonitrile to each well followed by 100 µL of serum. Mix thoroughly by repeated aspirate/dispense steps. Apply vacuum or pressure to elute the analytes.

Post Extraction

Dry the extract in a stream of air or nitrogen at 40 °C. Reconstitute with 100 µL of 0.4% formic acid (aq) and vortex for 30 seconds before analysis.

Summary of Results

Reproducible recovery (~85%) with low RSD (<2%). Extracts are protein free.

AN851

Extraction of Methylmalonic Acid from Serum Using ISOLUTE® SLE+ Prior to LC-MS/MS Analysis



Analytes

Methylmalonic acid (MMA).

Format

ISOLUTE® SLE+ 200 µL Supported Liquid Extraction plate, part number 820-0200-P01.



Matrices

Serum.

Sample Preparation Method

To serum (100 µL), add 10 µL of ISTD (10 ng/µL). Allow to equilibrate and add 4.6M formic acid (aq) (100 µL). Mix. Load the pre-treated serum (200 µL) onto the plate and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.

Elute analytes with MTBE (750 µL).

Post Extraction

Evaporate the extract in a stream of air or nitrogen at 40 °C. Addition of 2 µL of ethylene glycol prior to evaporation can help reduce analyte losses due to volatility. Reconstitute extracts with 100 µL of 0.4% formic acid (aq) before analysis.

Summary of Results

High reproducible recoveries >80% and corresponding RSDs of <10%.

AN850

Extraction of Methylmalonic Acid from Serum Using ISOLUTE® PLD+ Prior to LC-MS/MS Analysis

**Analytes**

Methylmalonic acid (MMA).

Format

ISOLUTE® PLD+ Protein and Phospholipid Removal plate, part number 918-0050-P01.

Matrices

Serum.

Sample Preparation Method

To serum (100 µL), add 10 µL of ISTD (10 ng/µL). Mix and allow to equilibrate. Place extraction plate in manifold, with an appropriate collection plate in position. Add 800 µL of 1% (v/v) formic acid in acetonitrile to each well followed by 100 µL of serum. Mix thoroughly by repeated aspirate/dispense steps. Apply vacuum or pressure to elute the analytes.

Post Extraction

Dry the extract in a stream of air or nitrogen at 40 °C. Reconstitute with 100 µL of 0.4% formic acid (aq) and vortex for 30 seconds before analysis.

Summary of Results

High reproducible recoveries >90% with corresponding RSDs <10%. Extracts are clean with no interference from phospholipids or proteins.

AN849

Extraction of Methylmalonic Acid from Serum Using ISOLUTE® SAX Prior to LC-MS/MS Analysis

**Analytes**

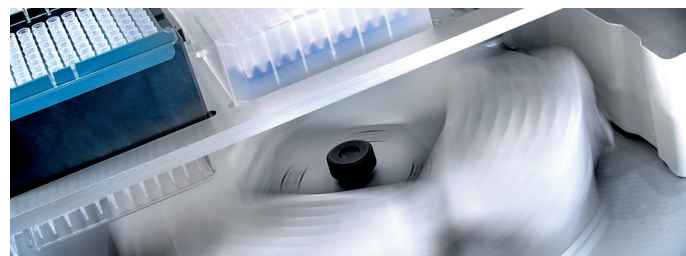
Methylmalonic acid (MMA).

Format

ISOLUTE® SAX 25 mg Fixed Well plate, part number 500-0025-P01.

Matrices

Serum.

**Sample Preparation Method**

To serum (100 µL), add 10 µL of internal standard (10 ng/µL). Mix and allow to equilibrate. Add HPLC grade water (190 µL) and vortex. Condition each well with methanol (500 µL) followed by HPLC grade water (500 µL). Load 300 µL of pre-treated sample. Elute interferences with HPLC grade water (500 µL) followed by methanol (500 µL). Elute analytes with 2% formic acid in acetonitrile (600 µL).

Post Extraction

Dry the extract in a stream of air or nitrogen at 40 °C. Reconstitute with 100 µL of 0.4% formic acid (aq) and vortex for 30 seconds before analysis.

Summary of Results

High reproducible recoveries >90% with corresponding RSDs <10%. Extracts are clean with no interference from phospholipids or proteins.

AN847

Extraction of Methylmalonic Acid from Serum Using EVOLUTE® EXPRESS AX Prior to LC-MS/MS Analysis



Analytes

Methylmalonic acid (MMA).

Format

EVOLUTE® EXPRESS AX 30 mg Fixed Well plate, part number 603-0030-PX01.



Matrices

Serum.

Sample Preparation Method

To serum (100 µL), add 10 µL of ISTD (10 ng/µL). Mix and allow to equilibrate. Add HPLC grade water (290 µL) and vortex. Load pre-treated sample (400 µL) direct to the 96-well plate. Elute interferences with HPLC grade water (1 mL) followed by methanol (1 mL). Elute analytes into a collection plate using 2% formic acid in acetonitrile (1 mL).

Post Extraction

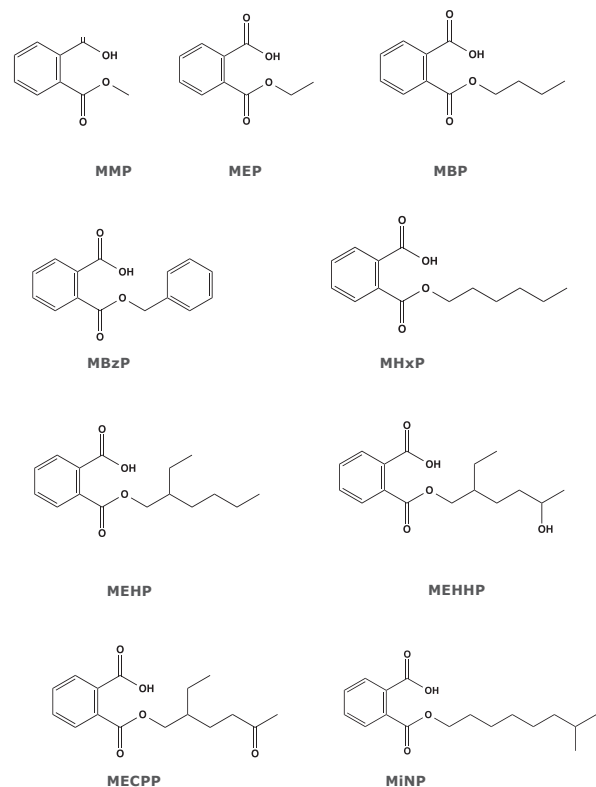
Dry the extract in a stream of air or nitrogen at 40 °C. Reconstitute with 100 µL of 0.4% formic acid (aq) and vortex for 30 seconds before analysis.

Summary of Results

High reproducible recoveries >90% and corresponding RSDs of <10%.

AN826

A High-Throughput SPE Method to Support the Biomonitoring of Phthalate Metabolites in Human Urine Using ISOLUTE® ENV+ Columns Prior to LC-MS/MS



Structures of the target analytes in the phthalate metabolites panel.

Analytes

Monomethyl phthalate (MMP); monoethyl phthalate (MEP); monobutyl phthalate (MBP); monobenzyl phthalate (MBzP); monohexyl phthalate (MHxP); mono(2-ethylhexyl) phthalate (MEHP); mono(2-ethyl-5 hydroxyhexyl) phthalate (MEHHP); mono(2-ethyl-5-carboxypentyl) phthalate (MECPP) and monoisooctyl phthalate (MiNP) .

Format

ISOLUTE® ENV+ 50 mg/3 mL columns, part number 915-0005-B.

Matrices

Urine.

Sample Preparation Method

Hydrolyze urine and add internal standard. Mix and allow to equilibrate. Condition columns with methanol (1 mL) and equilibrate with water (1 mL). Load 500 µL of pre-treated sample. Elute interferences with water/methanol (90/10, v/v, 1 mL), and elute analytes with methanol (2 x 1 mL).

Post Extraction

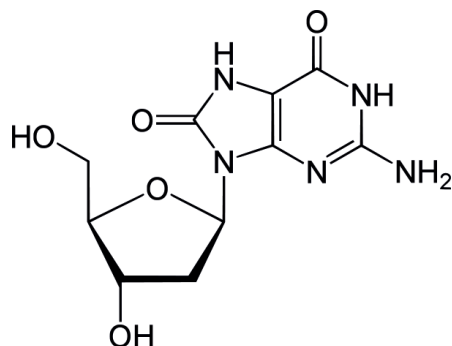
Evaporate to dryness and reconstitute before analysis.

Summary of Results

ISOLUTE® ENV+ SPE cartridges were successful in providing quantitative analyte recovery, repeatable method precision and minimal matrix suppression for nine phthalate metabolites in urine.

AN759

Extraction of 8-oxoDG from Biological Fluids Using ISOLUTE® ENV+



Structures of (8-oxo-DG).

Analytes

8-oxo-7,8-dihydro-2'-deoxyguanosine (8-oxo-DG).

Format

ISOLUTE® ENV+ 50 mg/1 mL columns, part number 915-0005-A.

Matrices

Urine, plasma, saliva, breast milk, seminal plasma, peritoneal fluid.

Sample Preparation Method

Urine: Centrifuge 0.15 mL of urine at 16,100 x g for 15 mins. Add internal standard and dilute 1:1 (v/v) with deionized water. [For all other matrices, see full application note]. Condition column with methanol (1 mL) and equilibrate with water (1 mL). Load sample (1 mL) and elute interferences with 2% (v/v) methanol (aq) (1 mL). Elute analyte with 20% (v/v) acetonitrile in methanol (2 x 300 µL).

Post Extraction

Evaporate to dryness and reconstitute in mobile phase (50 µL). Mix and centrifuge before analysis.

Summary of Results

Recovery >85% with low intra and inter day variability. LOQ in urine <1 pmol/mL.

AN737

Extraction of Organophosphate Pesticide Metabolites From Urine Using EVOLUTE® EXPRESS WAX

Analytes

Dimethylphosphate (DMP), dimethylthiophosphate (DMTP), diethylphosphate (DEP), diethylthiophosphate (DETP), dimethyldithiophosphate (DMDTP) and diethyldithiophosphate (DEDTP).

Format

EVOLUTE® EXPRESS WAX 30 mg Fixed Well plate, part number 604-0030-PX01.



Matrices

Urine

Sample Preparation Method

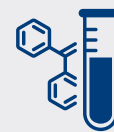
Dilute urine sample (100 µL) with 2 % formic acid (300 µL) (1:3, v/v). Condition column with methanol (1 mL) and equilibrate with HPLC grade water (1 mL). Load pre-treated sample (400 µL). Elute polar and ionic interferences with HPLC grade water (1 mL). Elute non-polar interferences with acetonitrile (1 mL). Elute analytes with methanol containing ammonium hydroxide (95:5, v/v, 1 mL).

Post Extraction

Evaporate to dryness and reconstitute in mobile phase before analysis.

Summary of Results

Typical recoveries for all analytes are above 85% with relative standard deviations below 10%.



Catecholamine Metabolites

AN874

Extraction of Plasma Catecholamines and Metanephrines Using EVOLUTE® EXPRESS WCX SPE Prior to LC-MS/MS Analysis



page 9

AN871

Extraction of Urinary Catecholamines and Metanephrines Using EVOLUTE® EXPRESS WCX SPE Prior to LC-MS/MS Analysis



page 10

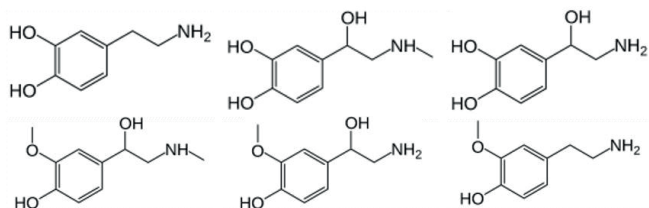


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AN874

Extraction of Plasma Catecholamines and Metanephrines Using EVOLUTE® EXPRESS WCX SPE Prior to LC-MS/MS Analysis



Structures of dopamine, epinephrine and norepinephrine (top), metanephrine, normetanephrine and 3-methoxytyramine (bottom).

Analytes

Epinephrine, norepinephrine, dopamine, metanephrine, normetanephrine and 3-methoxytyramine.

Format

EVOLUTE® EXPRESS WCX 10 mg Fixed Well plate, part number 602-0010-PX01.

Matrices

Plasma.

Sample Preparation Method

Mix plasma (300 µL) with 10 µL of internal standard solution and 0.05% formic acid (300 µL). Mix and allow to equilibrate. Load pre-treated plasma (500 µL). Elute interferences with 10 mM ammonium acetate (500 µL) followed by propan-2-ol (500 µL) and finally dichloromethane (500 µL). Elute analytes with 125 µL of water: propan-2-ol (85:15, v/v) containing formic acid (0.1% v/v).

Post Extraction

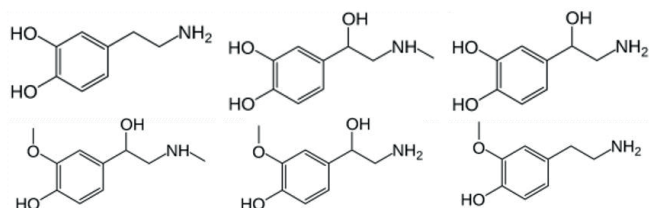
The extract can be injected directly into the analytical system without an additional evaporation step.

Summary of Results

High, reproducible analyte recoveries. Linearity was determined between 0.04 and 1.28 ng/mL for norepinephrine and dopamine and between 0.02 and 1.28 ng/mL for epinephrine, normetanephrine, metanephrine and 3-methoxytyramine.

AN871

Extraction of Urinary Catecholamines and Metanephrines Using EVOLUTE® EXPRESS WCX SPE Prior to LC-MS/MS Analysis



Structures of dopamine, epinephrine and norepinephrine (top), metanephrine, normetanephrine and 3-methoxytyramine (bottom).

Analytes

Epinephrine, norepinephrine, dopamine, metanephrine, normetanephrine and 3-methoxytyramine.

Format

EVOLUTE® EXPRESS WCX 10 mg Fixed Well plate, part number 602-0010-PX01.

Matrices

Urine

Sample Preparation Method

Mix urine (75 μ L) with 10 μ L of internal standard solution and 250 mM ammonium acetate solution (150 μ L). Mix and allow to equilibrate. ammonium acetate (500 μ L). Load pre-treated urine (150 μ L). Elute interferences with 10mM ammonium acetate (500 μ L) followed by propan-2-ol (500 μ L), and dry thoroughly. Elute analytes with 125 μ L of water: propan-2-ol (85:15, v/v) containing formic acid (0.1% v/v). Dry. Note that conditioning and equilibration steps are not required when using the EVOLUTE EXPRESS Load-Wash-Elute protocol.

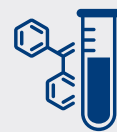
Post Extraction

The extract can be injected directly into the analytical system without an additional evaporation step.

Summary of Results

High reproducible recoveries with LOQs ranging from 0.1 ng/mL (epinephrine) to 2.5 ng/mL (dopamine).

Pain Management Drug Panel



PPS443	Extraction of a Urine Drug Panel Using ISOLUTE® SLE+	page 11
PPS443	Extraction of a Urine Drug Panel Using EVOLUTE® EXPRESS ABN	page 12
PPS443	Extraction of a Urine Drug Panel Using EVOLUTE® EXPRESS CX	page 12

PPS443

Extraction of a Urine Drug Panel Using ISOLUTE® SLE+

Analytes

56 drug panel—see list in Biotage White Paper PPS443.

Format

ISOLUTE® SLE+ 400 µL Supported Liquid Extraction plate, part number 820-0400-P01.

Matrices

Urine.

Sample Preparation Method

Hydrolyze urine (200 µL) using β glucuronidase enzyme (see PPS443 for hydrolysis conditions). Add 0.1% NH₄OH (200 µL) and mix. Load hydrolyzed sample (400 µL) and apply a pulse of pressure to initiate flow. Allow to absorb for 5 minutes.

Elute analytes with 90:10 (v/v) dichloromethane:2-propanol (DCM:IPA) (2 x 0.75 mL).

Post Extraction

Dry under nitrogen (N₂) at 40 °C. Reconstitute in 90:10 (v/v) 0.1% formic acid in water/0.1% formic acid in methanol before analysis.

Summary of Results

ISOLUTE® SLE+ is recommended if opiates, opioids, benzodiazepines, stimulants (except ritalinic acid), PCP, barbiturates, 9-carboxy-THC, TCAs, meprobamate, carisoprodol, ketamine and norketamine are included in the urine panel.



PPS443

Extraction of a Urine Drug Panel Using EVOLUTE® EXPRESS ABN

Analytes

56 drug panel—see list in Biotage White Paper PPS443.

Format

EVOLUTE® EXPRESS ABN 30 mg Fixed Well plate (600-0030-PX01).



Matrices

Urine.

Sample Preparation Method

Hydrolyze urine (200 µL) using β-glucuronidase enzyme (see PPS443 for hydrolysis conditions). Add 0.1% NH₄OH (200 µL) and mix. Condition wells with methanol (1 mL) and equilibrate with 0.1% NH₄OH (1 mL). Load hydrolyzed sample (400 µL). Elute interferences with 0.1% NH₄OH (1 mL) followed by 10 methanol in water (v/v, 1 mL). Elute analytes with DCM:IPA (90:10, v/v, 2 x 0.75 mL).

Post Extraction

Dry under nitrogen (N₂) at 40 °C. Reconstitute in 90:10 (v/v) 0.1% formic acid in water/0.1% formic acid in methanol before analysis.

Summary of Results

EVOLUTE® EXPRESS ABN works well for some opioid drugs and metabolites, most benzodiazepines, ketamine, norketamine, PCP, 9-carboxy-THC, amitriptyline, nortriptyline, carisoprodol, meprobamate, cocaine and BZE.

PPS443

Extraction of a Urine Drug Panel Using EVOLUTE® EXPRESS CX

Analytes

56 drug panel—see list in Biotage White Paper PPS443.

Format

EVOLUTE® EXPRESS CX 30 mg Fixed Well plate, part number 601-0030-PX01.

Matrices

Urine.

Sample Preparation Method

Hydrolyze urine (200 µL) using β-glucuronidase enzyme (see PPS443 for hydrolysis conditions). Add 4% phosphoric acid (H₃PO₄) (200 µL) and mix. Condition wells with methanol (1 mL) and equilibrate with 4% H₃PO₄ (1 mL). Load hydrolyzed sample (400 µL). Elute interferences with 4% H₃PO₄ (1 mL) followed by 50% methanol in water (v/v, 1 mL). Elute analytes with either:

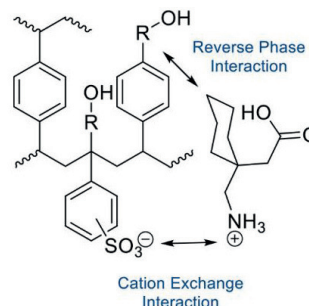
- 78:20:2 (v/v) DCM:IPA:NH₄OH (2 x 0.75 mL), or
- 78:20:2 (v/v) DCM:MeOH:NH₄OH (2 x 0.75 mL).

Post Extraction

Dry under nitrogen (N₂) at 40 °C. Reconstitute in 90:10 (v/v) 0.1% formic acid in water/0.1% formic acid in methanol before analysis.

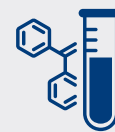
Summary of Results

If the drugs and metabolites in the urine panel are mostly basic: opiates, opioids, benzodiazepines, PCP, stimulants (except ritalinic acid), TCAs, ketamine, norketamine, and 9-carboxy-THC; then the EVOLUTE® EXPRESS CX method with sample pretreatment using 4% H₃PO₄, a 50% MeOH wash, and elution with 78:20:2 (v/v) DCM:IPA:NH₄OH is recommended. If ritalinic acid, gabapentin and pregabalin are required, the same protocol, except elution with 78:20:2 (v/v) DCM:MeOH:NH₄OH should be used.



EVOLUTE® EXPRESS CX sorbent's proposed columbic complexation with pregabalin.

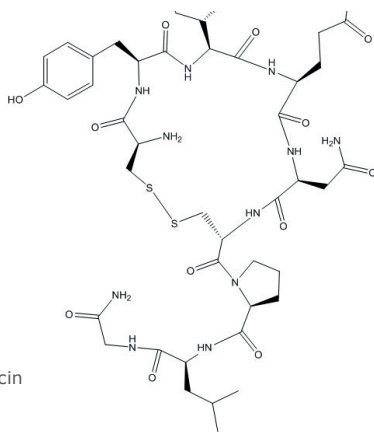
Peptides



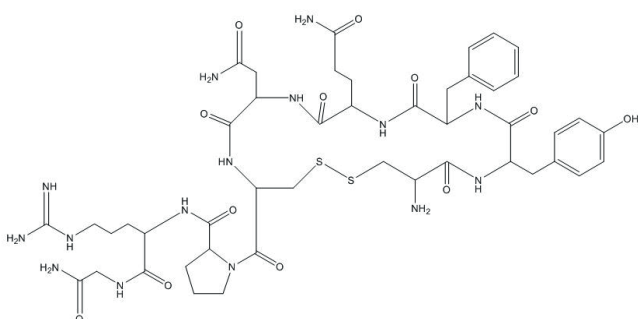
AN882	Extraction of Oxytocin and Vasopressin from Serum Using EVOLUTE® EXPRESS ABN Prior to LC-MS/MS Analysis	page 13
AN869	Extraction of Teicoplanin from Plasma Using EVOLUTE EXPRESS ABN Prior to HPLC-DAD Analysis	page 14

AN882

Extraction of Oxytocin and Vasopressin from Serum Using EVOLUTE® EXPRESS ABN Prior to LC-MS/MS Analysis



Oxytocin



Vasopressin

Analytes

Oxytocin and vasopressin.

Format

EVOLUTE® EXPRESS ABN 30 mg Fixed Well plate, part number 600-0030-PX01.

Matrices

Serum.

Sample Preparation Method

Dilute serum samples with 1% formic acid (1:1, v/v). Condition wells with methanol (1 mL) and equilibrate with 0.1% formic acid (aq) (1 mL). Load 400 µL of pre-treated serum sample. Elute interferences with 0.1% formic acid (aq) (1 mL). Elute analytes with 5% formic acid in acetonitrile /H₂O (20/80, v/v, 200 µL). *This highly aqueous elution solvent delivers high recoveries of oxytocin and vasopressin, and if desired, the extract can be injected directly into the analytical system without additional processing.* Note that conditioning and equilibration steps are not required when using the EVOLUTE® EXPRESS Load-Wash-Elute protocol.

Post Extraction

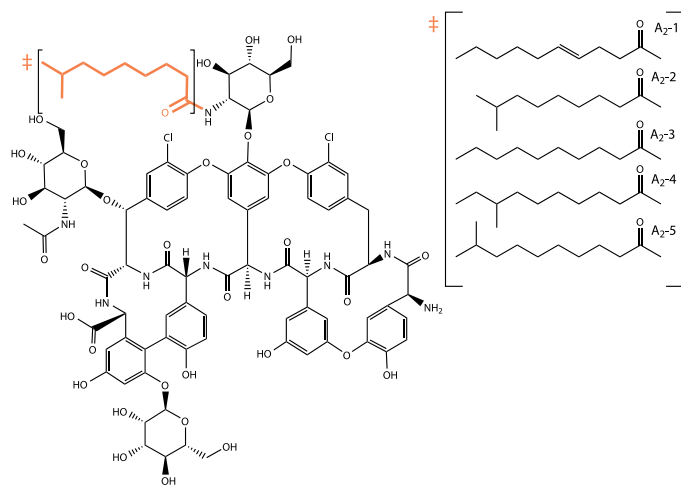
Alternatively, to minimize ion suppression, evaporate to dryness at 40 °C and reconstitute the extract with 0.1% formic acid in acetonitrile/H₂O (10/90, v/v, 200 µL) before analysis.

Summary of Results

High reproducible recoveries >70 % with corresponding RSDs <10%. Good sensitivity at concentrations down to 0.2 ng/mL.

AN869

Extraction of Teicoplanin from Plasma Using EVOLUTE® EXPRESS ABN Prior to HPLC-DAD Analysis



Teicoplanin A₂-2 (Major side-chain variants shown in parenthesis).

Analytes

Teicoplanin (as a mixture of A₂ variants).

Format

EVOLUTE® EXPRESS ABN 30 mg Fixed Well plate, part number 600-0030-PX01.

Matrices

Plasma.

Sample Preparation Method

Dilute plasma (200 µL) in a 1:3 ratio using 2% formic acid (aq). Condition each well with methanol (1 mL), and equilibrate with 0.1% formic acid (aq) (1 mL). Load 800 µL of pre-treated sample. Elute interferences with water (1 mL). Elute analytes with methanol/water (70/30, v/v, 500 µL).

Post Extraction

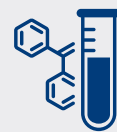
Evaporate to dryness at 40 °C and reconstitute with acetonitrile: 10 mM ammonium acetate pH 4.4 (10/90, v/v, 250 µL) before analysis.

Summary of Results

High recovery (88–100%) with low RSD (<7%). Linear range 2–100 µg/mL.



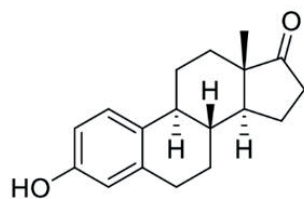
Steroids



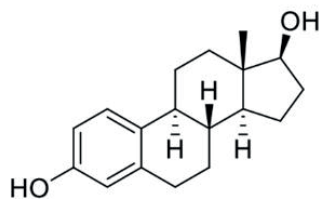
AN860	Extraction of Estrone and Estradiol from Human Serum Using ISOLUTE® SLE+ Prior to HPLC-MS/MS	page 16
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AN762	Extraction of Low Level Testosterone and Androstenedione from Human Serum Samples Using ISOLUTE® SLE+	page 17
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AN860

Extraction of Estrone and Estradiol from Human Serum Using ISOLUTE® SLE+ Prior to HPLC-MS/MS



Estrone (E1)
Molecular Weight: 270.37



Estradiol (E2)
Molecular Weight: 272.39

Structures of Estrone (E1) and Estradiol (E2).

Analytes

Estrone (E1) and estradiol (E2).

Format

ISOLUTE® SLE+ 400 µL Supported Liquid Extraction plate, part number 820-0400-P01.

Matrices

Serum

Sample Preparation Method

Dilute serum sample (250 µL) with 25% IPA (aq) (100 µL) and add IS. Mix and allow to equilibrate. Load 350 µL of sample onto the plate and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.

Elute analytes with dichloromethane (DCM) (3 x 600 µL).

Post Extraction

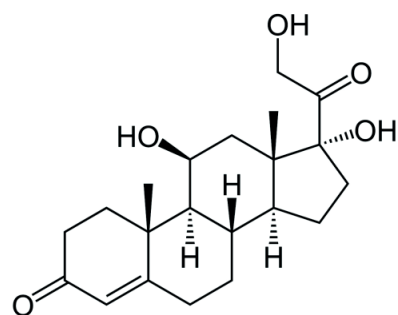
Evaporate to dryness at 40 °C and reconstitute with 250 µL of H₂O/ACN/MeOH (2:1:1, v/v) before analysis.

Summary of Results

High recovery and excellent sensitivity (LOQ of 0.001 ng/mL (estradiol) and 0.002 ng/mL (estrone) was achieved.

AN778

Extraction of Cortisol from Human Saliva Using ISOLUTE® SLE+ Plates Prior to LC-MS/MS Analysis



Structure of cortisol.

Analytes

Cortisol.

Format

ISOLUTE® SLE+ 200 µL Supported Liquid Extraction plate, part number 820-0200-P01.

Matrices

Oral fluid.

Sample Preparation Method

Dilute sample 1:1 (v/v) with water. Load the pre-treated sample (200 µL total volume) onto the plate and apply a pulse of vacuum or positive pressure to initiate flow. Allow the sample to adsorb for 5 minutes.

Elute analytes with MTBE (1 mL).

Post Extraction

Evaporate the extract to dryness. Reconstitute in water:methanol (50:50, v/v) (100 µL) before analysis.

Summary of Results

Recoveries greater than 95% with RSDs below 3%.

AN777

Extraction Cortisol from Human Urine Using ISOLUTE® SLE+ Plates Prior to LC-MS/MS Analysis

Analytes

Cortisol

Format

ISOLUTE® SLE+ 200 µL Supported Liquid Extraction plate, part number 820-0200-P01.



Matrices

Urine

Sample Preparation Method

Dilute sample 1:1 (v/v) with water. Load the pre-treated sample (200 µL total volume) onto the plate and apply a pulse of vacuum or positive pressure to initiate flow. Allow the sample to adsorb for 5 minutes.

Elute analytes with MTBE (1 mL).

Post Extraction

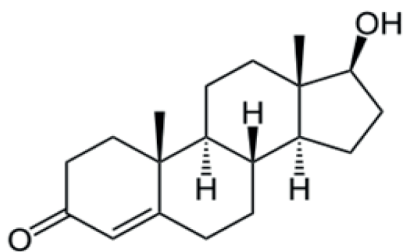
Evaporate the extract to dryness (40 °C). Reconstitute in water:methanol (50:50, v/v) (100 µL) before analysis.

Summary of Results

Recoveries greater than 99% with RSDs below 5%.

AN762

Extraction of Low Level Testosterone and Androstenedione from Human Serum Samples Using ISOLUTE® SLE+



Structure of testosterone.

Analytes

Testosterone, androstenedione.

Format

ISOLUTE® SLE+ 400 µL Supported Liquid Extraction plate, part number 820-0400-P01.

Matrices

Serum

Sample Preparation Method

Dilute human serum (200 µL) with 0.5 mol/L ammonium hydroxide (200 µL), add internal standard and mix.

Load the pre-treated sample (400 µL) on to the plate and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.

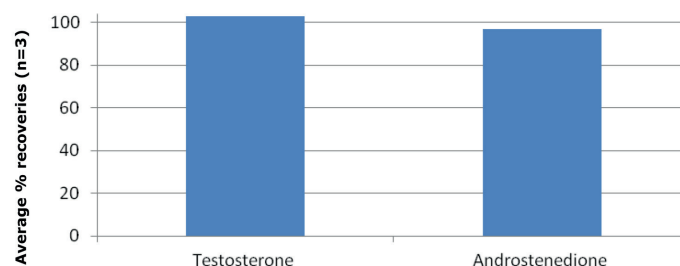
Elute analytes with diethyl ether (3 x 500 µL).

Post Extraction

Evaporate the eluate to dryness and reconstitute with 400 µL of methanol:water (1:1, v/v) before analysis.

Additional information: testosterone has an affinity to bind to plastic so the extracts were collected in glass tubes held in a 96 well collection plate.

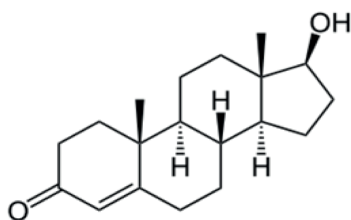
Summary of Results



Average analyte recoveries up to 100 nmol/L.

AN740

Extraction of Testosterone and Other Steroid Hormones from Human Plasma Using ISOLUTE® SLE+ 96-Well Plates



Structure of testosterone.

Analytes

Testosterone, aldosterone, 21-deoxycortisol, 11-deoxycortisol, androstendione, 17- α -hydroxyprogesterone, dehydroepiandrosterone (DHEA), progesterone, androsterone.

Format

ISOLUTE® SLE+ 200 μ L Supported Liquid Extraction plate, part number 820-0200-P01.

Matrices

Human plasma

Sample Preparation Method

Dilute human plasma (100 μ L) 1:1 with HPLC grade water (100 μ L). Load the pre-treated sample (200 μ L) on to the plate and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.

Elute analytes with dichloromethane (1 mL).

Post Extraction

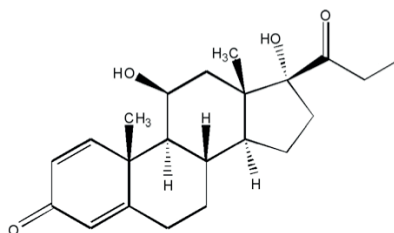
Evaporate to dryness at ambient temperature and reconstitute in 50% methanol (aq) (100 μ L). Vortex samples to ensure full reconstitution of all analytes.

Summary of Results

Recoveries ranging from 90–107% ($n=7$). The LOD for each analyte ranging from 0.5 ng/mL for androstendione to 100 ng/mL for DHEA.

AN602

Extraction of Corticosteroids from Plasma Using ISOLUTE® SLE+ Supported Liquid Extraction Plates



Structure of prednisone.

Analytes

Triamcinolone, prednisolone, hydrocortisone, prednisone, cortisone, betamethasone, dexamethasone, flumethasone, corticosterone, beclomethasone, triamcinolone acetonide, fluocinolone acetonide, budesonide structural isomer 1, budesonide structural isomer 2, 5-pregnen-3 β -ol-20-one.

Format

ISOLUTE® SLE+ 200 μ L Supported Liquid Extraction plate, part number 820-0200-P01.

Matrices

Plasma.

Sample Preparation Method

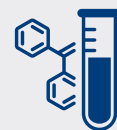
Dilute the sample with water (1:1, v/v) and mix. Load the pre-treated plasma (200 μ L) onto the plate, and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.

Post Extraction

Evaporate the extracts to dryness and reconstitute in H₂O/MeOH (80:20, v/v, 500 μ L) before analysis.

Summary of Results

Analyte	% Recovery	RSD (%)
Triamcinolone	93	4
Prednisolone	93	1
Hydrocortisone	98	2
Prednisone	95	1
Cortisone	96	2
Betamethasone	92	2
Dexamethasone	92	2
Flumethasone	91	2
Corticosterone	94	2
Beclomethasone	91	2
Triamcinolone Acetonide	91	3
Fluocinolone Acetonide	90	3
Budesonide Structural Isomer 1	87	3
Budesonide Structural Isomer 2	89	2
5-pregnen-3 β -ol-20-one	95	4

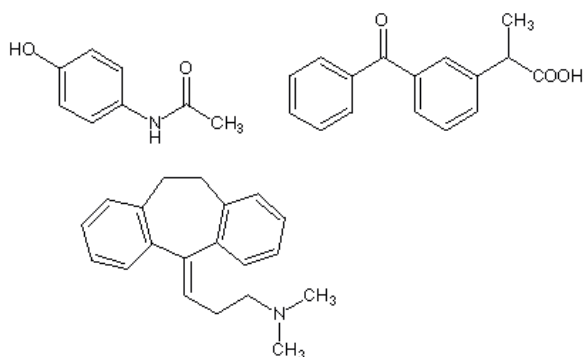


Therapeutic Drugs

AN830	Extraction of a Range of Acidic, Basic and Neutral Drugs from Plasma Using ISOLUTE® PLD+ Plates Prior to LC-MS/MS Analysis	page 20
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AN830

Extraction of a Range of Acidic, Basic and Neutral Drugs from Plasma Using ISOLUTE® PLD+ Plates Prior to LC-MS/MS Analysis



Structures of acetaminophen (neutral), ketoprofen (acidic) and amitriptyline (basic): examples of the broad range of analytes extracted in this application.

Analytes

Acetaminophen, amitriptyline, atenolol, bretylium tosylate, brompheniramine, fluoxetine, metoprolol, mianserin, naltrexone, procainamide, quinidine, ranitidine, salbutamol, sulindac, p-toluamide and ketoprofen.

Format

ISOLUTE® PLD+ Protein and Phospholipid Removal Plate, part number 918-0050-P01.

Matrices

Plasma

Sample Preparation Method

Add internal standard to the plasma sample, mix, and allow to equilibrate. Place the extraction plate in manifold, with an appropriate collection plate in position. Add 400 µL of acetonitrile to each well followed by 100 µL of plasma. Mix thoroughly by repeated aspirate/dispense steps. Apply vacuum or pressure to elute the analytes.

Post Extraction

Evaporate to dryness at 40 °C. Reconstitute in 0.1% formic acid aq/methanol (80/20, v/v, 200 µL) before analysis.

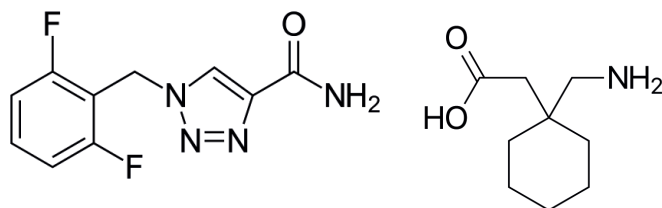
Summary of Results

A simple, generic approach to extraction of a broad range of analytes, giving high recoveries with low RSDs.



AN811

Extraction of Antiepileptic Drugs from Oral Fluid Using ISOLUTE® SLE+ Prior to LC-MS/MS Analysis



Structures of rufinamide and gabapentin.

Analytes

Tiagabine, carbamazepine-10,11-epoxide, oxcarbazepine, gabapentin, vigabatrin, rufinamide, felbamate.

Format

ISOLUTE® SLE+ 400 µL Supported Liquid Extraction plate, part number 820-0400-P01.

Matrices

Oral fluid.

Sample Preparation Method

Add ammonium acetate (5mM, pH 2.9, 250µL) to the sample (100 µL) then add up to 50 µL of internal standard. Mix. Load up to 400 µL of pre-treated oral fluid sample onto the plate and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes. Elute analytes with methyl tert-butyl ether containing 1% (v/v) trifluoroacetic acid (conc) solution (2 x 700 µL).

Post Extraction

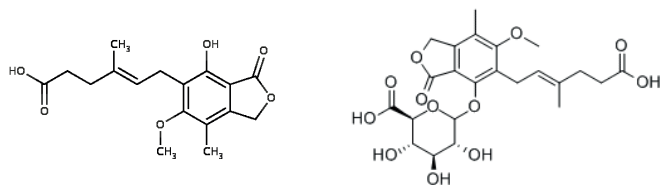
Evaporate to dryness and reconstitute sample with mobile phase before analysis.

Summary of Results

Recoveries for the AEDs using this pre-treatment strategy were good for all of the neutral AEDs in either neat or buffered oral fluid and substantially lower for the zwitterionic AEDs. Recovery >80% for felamate, rufinamide, oxcarbazepine, tiagabine and carbamazepine epoxide. Recoveries for gabapentin and vigabatrin were lower and attributed to their zwitterionic characteristics.

AN810

Extraction of Mycophenolic Acid (MPA) and Mycophenolic Acid Glucuronide (MPAG) from Serum Using ISOLUTE® SLE+ Prior to LC-MS/MS



Structure of mycophenolic acid (MPA) and mycophenolic acid glucuronide (MPAG).

Analytes

Mycophenolic acid (MPA) and mycophenolic acid glucuronide (MPAG).

Format

ISOLUTE® SLE+ 200 µL Supported Liquid Extraction plate, part number 820-0200-P01.

Matrices

Serum.

Sample Preparation Method

Add 90 µL of 20% aqueous formic acid to the sample (100 µL) then gently mix. Load the pre-treated sample onto the plate and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes. Elute analytes with ethyl acetate (2 x 500 µL).

Post Extraction

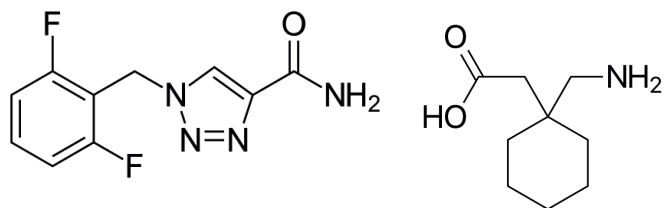
Evaporate to dryness and reconstitute in water:acetonitrile (50:50, v/v, 500 µL) before analysis.

Summary of Results

Recoveries >70% with RSDs <10%.

AN805

Extraction of Antiepileptic Drugs from Serum and Urine Using ISOLUTE® SLE+ Prior to LC-MS/MS Analysis



Structures of rufinamide and gabapentin.

Analytes

Tiagabine, carbamazepine-10,11-epoxide, oxcarbazepine, gabapentin, vigabatrin, rufinamide, felbamate.

Format

ISOLUTE® SLE+ 400 µL Supported Liquid Extraction plate, part number 820-0400-PO1.

Matrices

Serum, urine.

Sample Preparation Method

Neutral Antiepileptic Drugs in Serum and Urine:

Add ammonium acetate (5 mM, pH 2.9, 250 µL) to the sample (100 µL) then add up to 50 µL of internal standard and mix.

Load up to 400 µL of pre-treated serum/urine sample onto the plate. Apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.

Elute analytes with methyl tert-butyl ether containing 1% (v/v) trifluoroacetic acid (conc) solution (2 x 700 µL).

Neutral Antiepileptic and Zwitterionic Drugs in Serum and Urine:

Add 50% aqueous formic acid (100 µL) to the sample (100 µL) then add up to 100 µL of internal standard and mix.

Load up to 300 µL of the pre-treated serum sample onto the plate. Apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.

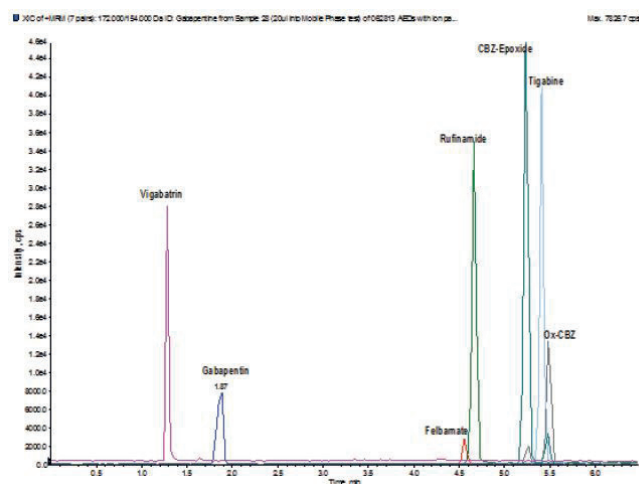
Elute analytes with methyl tert-butyl ether containing 1% (v/v) trifluoroacetic acid (conc) solution (2 x 700 µL).

Post Extraction

Evaporate to dryness at 40 °C and reconstitute sample in mobile phase before analysis.

Summary of Results

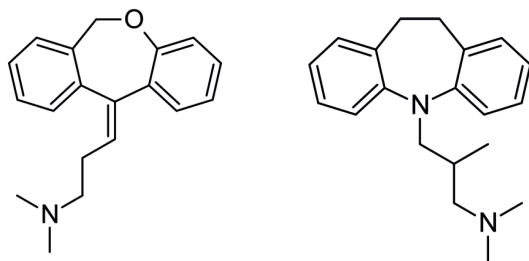
Recoveries >70% with RSD <10%.



Extracted ion chromatogram of antiepileptic drugs.

AN760

Fast Extraction of 10 Tricyclic Anti-depressant Drugs from Urine Using ISOLUTE® SLE+ Columns Prior to LC-MS-MS Analysis



Structures of doxepin and trimipramine.

Analytes

Trimipramine, imipramine, desipramine, clomipramine, amitriptyline, doxepin, desmethyldoxepin, nortriptyline, paroxetine, sertraline.

Format

ISOLUTE® SLE+ 200 µL Supported Liquid Extraction columns, part number 820-0140-C.

Matrices

Urine.

Sample Preparation Method

Mix urine with concentrated ammonium hydroxide (99:1, v/v). Add internal standard and mix. Load the pre-treated sample (1 mL) onto cartridge. Apply a short pulse of vacuum or positive pressure to initiate flow and allow sample to adsorb for 5 minutes.

Elute analytes with hexane:isopropanol (98:2, v/v, 2 x 4 mL).

Post Extraction

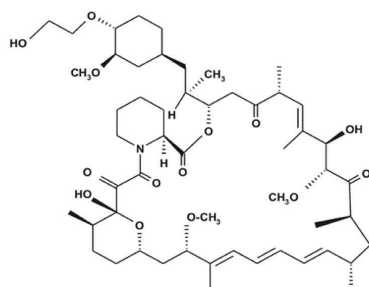
Evaporate sample to dryness and reconstitute in mobile phase (500 µL) before analysis.

Summary of Results

Recoveries for all of the TCAs were found to be >85% except for paroxetine which was observed at a recovery of 75%.

AN758

Extraction of a Range of Immunosuppressants from Whole Blood Using ISOLUTE® SLE+ for LC-MS/MS Analysis



Structure of everolimus.

Analytes

Sirolimus, tacrolimus, everolimus, cyclosporin A.

Format

ISOLUTE® SLE+ 400 µL Supported Liquid Extraction plate, part number 820-0400-P01.

Matrices

Whole blood.

Sample Preparation Method

In a 2 mL Eppendorf centrifuge tube, pipette whole blood (50 µL). Add HPLC water (250 µL) and vortex for 30 seconds. Centrifuge at 12,000 RPM for 10 minutes.

Load the supernatant (275 µL) onto the plate and apply a pulse of vacuum or positive pressure for 10 seconds. Allow the sample to absorb for 5 minutes.

Elute analytes with ethyl acetate (2 x 600 µL).

Post Extraction

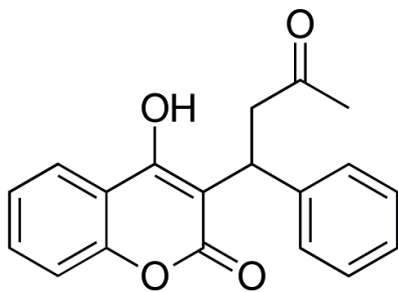
Evaporate the extract to dryness (30 °C). Reconstitute in water:acetonitrile (100 µL, 25:75, v/v).

Summary of Results

Recoveries ranged from 60–97%. RSDs were all below 10% for all analytes.

AN734

Method for the Extraction of Warfarin From Human Plasma Using ISOLUTE® SLE+



Structure of warfarin.

Analytes

Warfarin.

Format

ISOLUTE® SLE+ 200 µL Supported Liquid Extraction plate, part number 820-0200-P01.

Matrices

Human plasma.

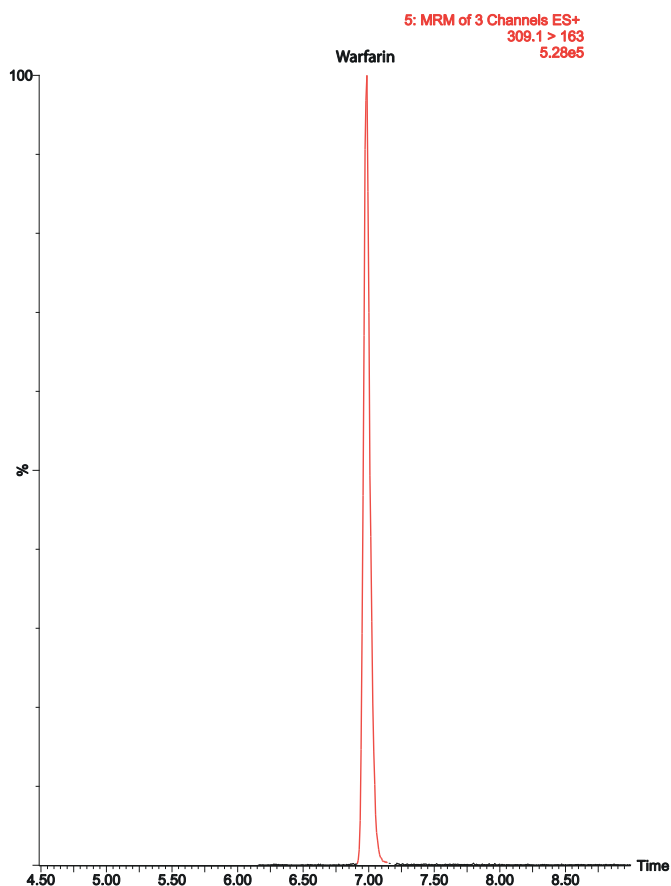
Sample Preparation Method

To plasma (100 µL) add 1% formic acid (100 µL), and mix. Load the pre-treated plasma (200 µL) onto the plate and allow to absorb for 5 mins.

Elute analytes with dichloromethane (DCM) (1 mL).

Summary of Results

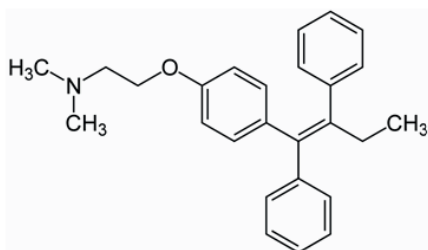
All results show recoveries above 90% with RSDs below 10%.



Example chromatogram showing warfarin.

AN721

Extraction of Tamoxifen and Metabolites from Urine Using ISOLUTE® SLE+



Structure of tamoxifen.

Analytes

Tamoxifen, endoxifen, 4-OH-tamoxifen, des-methyl-tamoxifen.

Format

ISOLUTE® SLE+ 200 µL Supported Liquid Extraction plate, part number 820-0200-P01.

Matrices

Urine.

Sample Preparation Method

Dilute 100 µL of urine 1:1 (v/v) with 0.5 M NH₄OH. Load the pre-treated urine (200 µL) onto the plate and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.

Elute analytes with ethyl acetate (2 x 500 µL).

Post Extraction

Evaporate to dryness and reconstitute in 500 µL of 0.1% formic acid in H₂O/MeOH (50:50, v/v) before analysis.

Summary of Results

All results show recoveries above 80% with %RSDs below 10%.

AN700

Extraction of Diuretics from Urine Using EVOLUTE® EXPRESS ABN Columns

Analytes

Amiloride, acetazolamide, hydrochlorothiazide, methazolamide, hydroflumethiazide, furosemide, bendoflumethiazide, bumetanide, spironolactone, ethacrynic acid.

Format

EVOLUTE® EXPRESS ABN 100 mg/3 mL columns (tablets), part number 610-0010-BXG.

Matrices

Urine.

Sample Preparation Method

Dilute urine samples with acetonitrile (1:9, v/v) and add internal standard as required. Mix. Condition columns with methanol (3 mL) and equilibrate with water (3 mL) followed by acetonitrile (3 mL). Load the pre-treated sample (2 mL). Elute interferences with acetonitrile (3 mL) followed by methanol (3 mL). Elute analytes with 2% HCl in acetonitrile (3 mL).

Post Extraction

Evaporate to dryness and reconstitute in H₂O/MeOH (50:50, v/v, 1 mL) for before analysis.

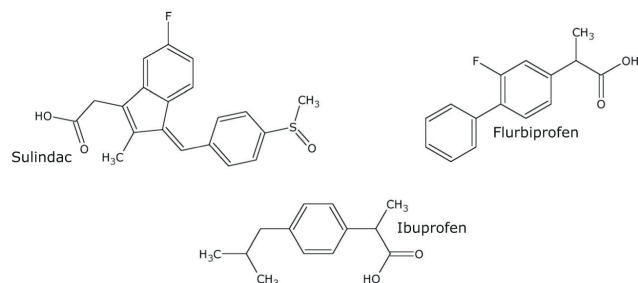
Summary of Results

High analyte recoveries (80–100%) with RSDs of <10% were achieved.



AN603

Extraction of Non-steroidal Anti-inflammatory Drugs (NSAIDs) from Plasma Using ISOLUTE® SLE+ Supported Liquid Extraction Plates



Structures of sulindac, flurbiprofen and ibuprofen.

Analytes

Sulindac, flurbiprofen, ibuprofen.

Format

ISOLUTE® SLE+ 400 µL Supported Liquid Extraction plate, part number 820-0400-P01.

Matrices

Plasma.

Sample Preparation Method

Dilute plasma (1:1, v/v) with 1% formic acid and mix. Load the pre-treated sample (200 µL) onto the plate and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.

Elute analytes with MTBE (2 x 900 µL).

Post Extraction

Evaporate to dryness and reconstitute in H₂O/MeOH (60:40, v/v, 500 µL).

Summary of Results

Analyte	Recovery	RSDs
Sulindac	92	3
Flurbiprofen	94	3
Ibuprofen	91	10

AN601

Extraction of Tricyclic Anti-depressants from Plasma Using ISOLUTE® SLE+ Supported Liquid Extraction Plates

Analytes

Imipramine, trimipramine, nortriptyline.

Format

ISOLUTE® SLE+ 200 µL Supported Liquid Extraction plate, part number 820-0200-P01.

Matrices

Plasma.

Sample Preparation Method

Dilute the sample (100 µL) with 0.5 M NH₄OH (100 µL). Mix. Load the pre-treated sample onto the plate and apply a pulse of vacuum or positive pressure to initiate flow. Allow to absorb for 5 minutes.

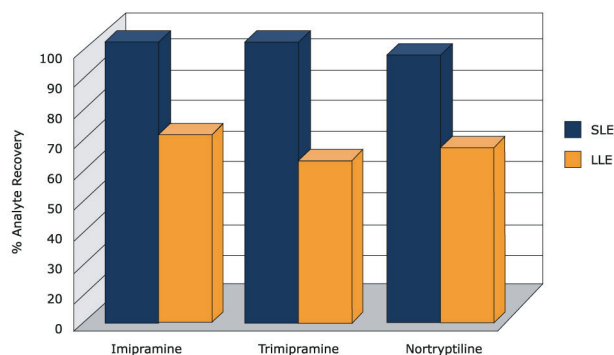
Elute analytes with hexane:3-methyl-1-butanol (98:2, v/v, 1 mL).

Post Extraction

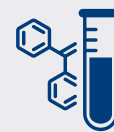
Evaporate to dryness and reconstitute in mobile phase (H₂O:ACN:NH₄OH 10:90:0.1, v/v) before analysis.

Summary of Results

Analyte recoveries were >91% with RDS %<4 for all analytes.



Comparison of analyte recovery using ISOLUTE® SLE+ and LLE.



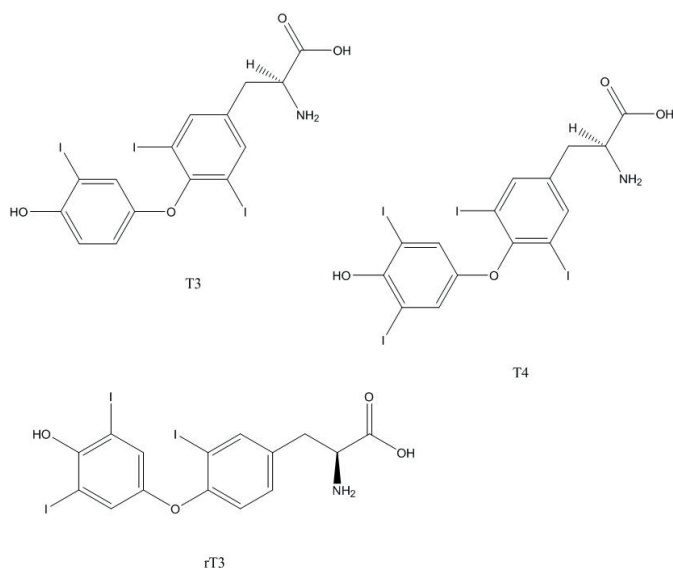
Thyroid Hormones

AN881 Extraction of Thyroid Hormones: T₃, rT₃ and T₄ from Serum Using EVOLUTE® EXPRESS AX Plates Prior to LC-MS/MS

page 27

AN881

Extraction of Thyroid Hormones: T₃, rT₃ and T₄ from Serum Using EVOLUTE® EXPRESS AX Plates Prior to LC-MS/MS



Structures of T₃, rT₃ and T₄.

Analytes

Tri-iodothyronine (T₃), reverse tri-iodothyronine (rT₃) and thyroxine (T₄).

Format

EVOLUTE® EXPRESS AX 30 mg Fixed Well Plate, part number 603-0030-PX01.

Matrices

Serum.

Sample Preparation Method

To serum (200 µL) add internal standard solution (10 µL), then add a further 100 µL of a mixture of citric acid, ascorbic acid and DL-dithiothreitol (25 mg/mL), and vortex mix thoroughly.

Note: the use of these stabilizers prevents conversion of T₄ to T₃ and rT₃ during extraction.

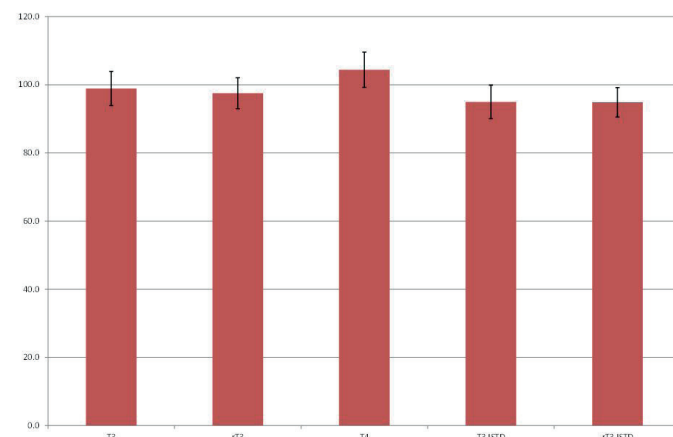
Condition each well with methanol (1 mL) and equilibrate with water (1 mL). Load the entire sample volume. Elute interferences with 50 mM NH₄OAc buffer, pH 9 aq (1 mL) followed by MeOH (1 mL), and finally 2% formic acid in DCM (v/v) (1 mL). Elute analytes with MeOH (500 µL).

Post Extraction

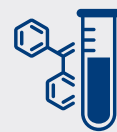
Evaporate to dryness at 40 °C and reconstitute with H₂O/MeOH (50/50 (v/v), 150 µL) before analysis.

Summary of Results

High reproducible recoveries >85% with corresponding RSDs <10%. Extremely clean extracts.



Recovery profile for thyroid hormones extracted at 2 ng/mL.

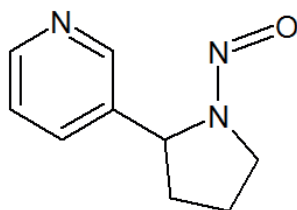


Tobacco Exposure

AN884	Extraction of Tobacco-Specific Nitrosamines (TSNAs) from Urine Using ISOLUTE® SLE+ Prior to UPLC/MS/MS Analysis	page 28
AN787	Extraction of Nicotine and Metabolites from Urine, Serum, Plasma and Whole Blood Using ISOLUTE® SLE+ Prior to LC-MS/MS Analysis	page 29

AN884

Extraction of Tobacco-Specific Nitrosamines (TSNAs) from Urine Using ISOLUTE® SLE+ Prior to UPLC/MS/MS Analysis



Structure of N-nitrosornicotine (NNN).

Analytes

NNN (n-nitrosornicotine), also suitable for NNK (4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone) and NNAL (4-(methylnitrosamino)-1-(3-pyridyl)-1-butanol).

Format

ISOLUTE® SLE+ 1 mL sample volume columns (tablets), part number 820-00140-CG.

Matrices

Urine

Sample Preparation Method

Load 1 mL of the IS spiked urine sample, and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.

Elute analytes with dichloromethane (DCM) (2 x 1.5 mL).

Post Extraction

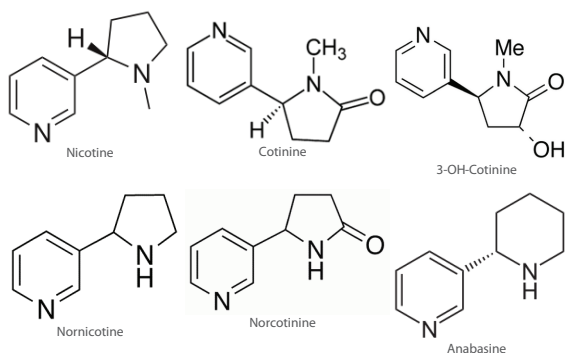
Dry under nitrogen (N₂) at 40 °C. Reconstitute in 0.1% formic acid in water/0.1% formic acid in methanol (90:10, v/v) before analysis.

Summary of Results

Limits of quantitation of 10 pg/mL can be achieved.

AN787

Extraction of Nicotine and Metabolites from Urine, Serum, Plasma and Whole Blood Using ISOLUTE® SLE+ Prior to LC-MS/MS Analysis



Structures of nicotine and metabolites.

Analytes

Nicotine, cotinine, 3-OH-cotinine, nor nicotine, norcotinine, anabasine.

Format

ISOLUTE® SLE+ 200 µL Supported Liquid Extraction plate, part number 820-0200-PO1.

Matrices

Urine, serum, plasma and whole blood.

Sample Preparation Method

To sample matrix (120 µL) add 10 µL internal standard and ammonia solution (25%, 230 µL). Mix well. Load pre-treated sample (150 µL) onto each well. Apply a pulse of vacuum or positive pressure to 3–5 secs.) to initiate flow. Allow the sample to absorb for 5 minutes.

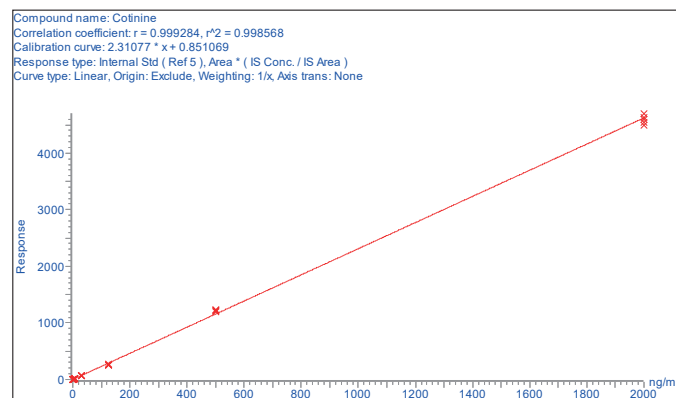
Elute analytes with dichloromethane:isopropanol (95: 5, v/v, 1 mL) into a 2 mL deep well collection plate containing 100 µL methanolic 200 mM HCl in each well.

Post Extraction

Dry the eluate in a stream of air or nitrogen. Reconstitute in methanol:water (10:90, v/v, 200 µL).

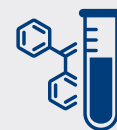
Summary of Results

Matrix	Recovery (%) of nicotine	RSD (%)
Urine	97.1	<7
Plasma	95.7	<5
Serum	95.5	<3
Whole Blood	97.9	<4



Shows typical linearity data achieved using this method.





Vitamins

AN880	Extraction of Vitamin B7 (Biotin) from Serum Using EVOLUTE® EXPRESS ABN Prior to LC-MS/MS Analysis		page 31
AN857	Ultra-Sensitive Method for the Determination of 1,25 di-OH Vitamin D2 and 1,25 di-OH Vitamin D3 in Serum Using Supported Liquid Extraction Prior to LC-MS/MS		page 31
AN842	Extraction of 25-hydroxy Vitamin D from Serum Using ISOLUTE® PLD+ Prior to LC-MS/MS Analysis		page 32
AN814	A High-Throughput SPE Method for Extraction of Vitamin B3 (Niacin) and Related Metabolites from Serum Using ISOLUTE® SCX-3 Prior to LC-MS/MS		page 32
AN757	Extraction of Vitamin D Metabolites from Human Serum Using ISOLUTE® SLE+ in 96-Fixed Well Plate Format Prior to LC-MS-MS Analysis		page 33
AN753	Extraction of Retinol, β -Carotene (Vitamin A) and α -Tocopherol (Vitamin E) from Serum Using ISOLUTE® SLE+ 96-Well Plates with APCI-LC-MS-MS Analysis		page 34

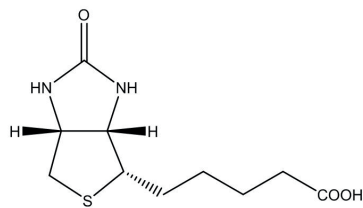


Automated Using Biotage® Extrahera™

Methods marked with this icon have been automated using Biotage® Extrahera™, and detailed settings are available.

AN88o

Extraction of Vitamin B7 (Biotin) from Serum Using EVOLUTE® EXPRESS ABN Prior to LC-MS/MS Analysis



Structure of Vitamin B7.

Analytes

25-OH vitamin D2 and 25-OH vitamin D3.

Format

EVOLUTE® EXPRESS ABN 10 mg Fixed Well plate, part number 600-0010-PX01.

Matrices

Serum.

Sample Preparation Method

Dilute serum (200 µL) with 1% formic acid (aq) (200 µL), add internal standard and mix. Condition wells with methanol (500 µL) and equilibrate with 1% formic acid (aq) (500 µL). Load 400 µL of diluted sample. Elute interferences with H₂O (500 µL) followed by H₂O/MeOH (95/5, v/v, 500 µL). Elute biotin with 0.1% NH₄OH in H₂O/MeOH (90/10, v/v, 200 µL). Note that conditioning and equilibration steps are not required when using the EVOLUTE® EXPRESS Load-Wash-Elute protocol.

Post Extraction

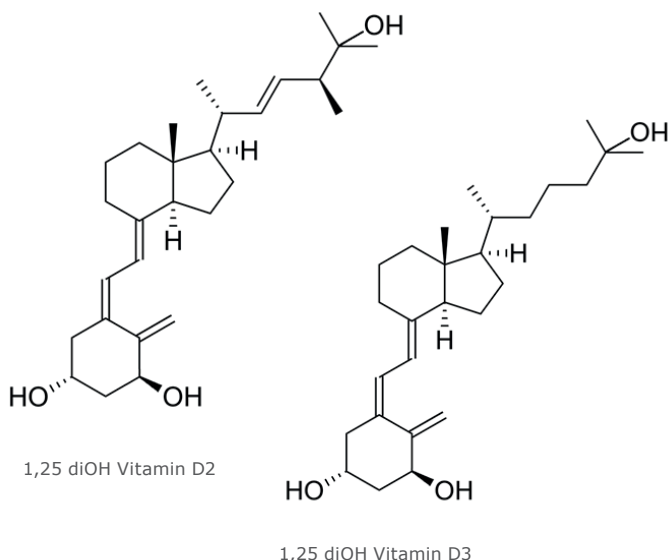
Evaporate extract to dryness at 40 °C in a stream of air or nitrogen. Reconstitute the extract with H₂O/ACN (90/10, v/v, 200 µL) before analysis.

Summary of Results

High reproducible recoveries >80% with corresponding RSDs <10%. Extracts are clean with minimal matrix effects.

AN857

Ultra-Sensitive Method for the Determination of 1,25 di-OH Vitamin D2 and 1,25 di-OH Vitamin D3 in Serum Using Supported Liquid Extraction Prior to LC-MS/MS



Analyte structures.

Analytes

1,25 diOH Vitamin D2 and 1,25 diOH Vitamin D3.

Format

ISOLUTE® SLE+ 400 µL Supported Liquid Extraction plate, part number 820-0400-P01.

Matrices

Serum.

Sample Preparation Method

To serum sample add internal standard, mix and leave to stand for at least 30 mins. Dilute the serum sample with an equal volume of volume of propan-2-ol : water (50:50, v/v) solution, mix. Load pre-treated serum (300 µL) and apply a pulse of vacuum or positive pressure (3–5 sec) to initiate flow. Allow the sample to absorb for 5 minutes.

Ensure a collection plate containing 200 µL of derivatization solution (0.25 mg/mL PTAD in ethyl acetate : heptane (8:92, v/v)) in each well is in position. Elute analytes with heptane (2 x 700 µL).

Post Extraction

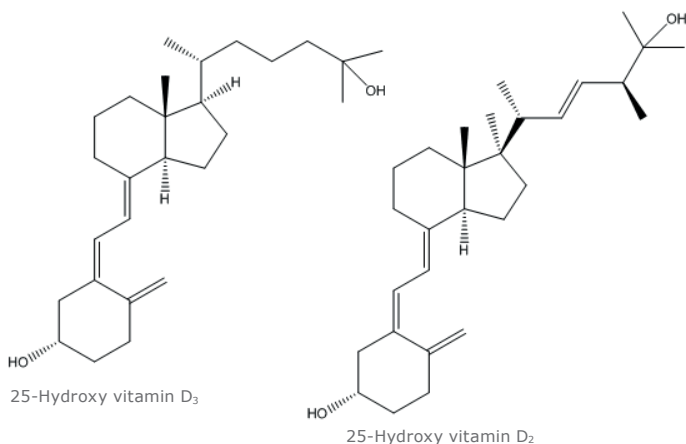
Evaporate the extracts to dryness at 40 °C and reconstitute in 70% methanol (aq) containing 50 µL/L methylamine (150 µL) before analysis.

Summary of Results

High reproducible recoveries are achieved. Linearity demonstrated from 5 to 500 pg/mL.

AN842

Extraction of 25-hydroxy Vitamin D from Serum Using ISOLUTE® PLD+ Prior to LC-MS/MS Analysis



Structures of 25-hydroxy Vitamin D.

Analytes

25-OH vitamin D₂ and 25-OH vitamin D₃.

Format

ISOLUTE® PLD+ Protein and Phospholipid Removal plate, part number 918-0050-P01.

Matrices

Serum.

Sample Preparation Method

To serum add internal standard, mix and leave to stand for at least 30 mins. Ensure a suitable collection plate is in position. Apply 400 µL of acetonitrile (MeCN) to each well of the ISOLUTE® PLD+ plate. Add 100 µL of serum with ISTD and mix thoroughly via repeat aspirate/dispense steps. Apply vacuum (-0.2 bar) or 3 psi positive pressure for approximately 5 minutes. For highly particulate laden samples increased pressure or vacuum conditions may be required.

Post Extraction

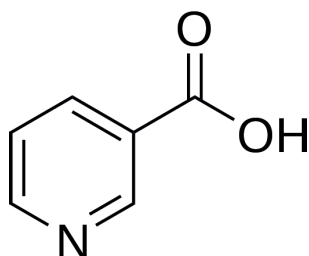
Dry the extract at 40 °C. Reconstitute using 100 µL of (30/70, v/v) 2 mM ammonium formate, 0.1% formic acid aq/MeOH.

Summary of Results

High analyte recovery with low RSDs was achieved. Linearity was demonstrated from 1–100 ng/mL. Five DEQAS samples were tested using this method and all determined values met acceptability criteria.

AN814

A High-Throughput SPE Method for Extraction of Vitamin B3 (Niacin) and Related Metabolites from Serum Using ISOLUTE® SCX-3 Prior to LC-MS/MS



Structure of Niacin (vitamin B₃).

Analytes

Niacin (nicotinic acid), nicotinuric acid, niacinamide.

Format

ISOLUTE® SCX-3 25 mg Fixed Well plate, part number 533-0025-P01.

Matrices

Serum.

Sample Preparation Method

Dilute serum (50 µL) with aqueous acetic acid (2%, 150 µL). Mix thoroughly. Condition each well with methanol (1 mL) and equilibrate with aqueous acetic acid (2%, 1 mL). Load pre-treated sample (200 µL). Wash each well with water:methanol:acetic acid (68:30:2, v/v/v, 2 x 1 mL) followed by methanol:acetic acid (98: 2, v/v, 2 x 1 mL). Elute analytes with methanol: ammonium hydroxide (95:5, v/v, 2 x 400 µL).

Post Extraction

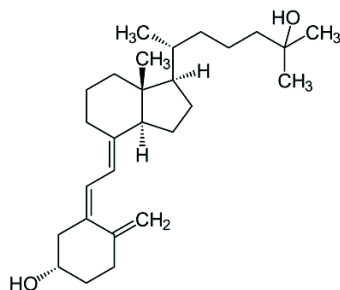
Evaporate extracts to dryness and reconstitute in 0.1% formic acid (100 µL) prior to analysis.

Summary of Results

This method is a viable option for serum measurements over a relevant concentration range in clinical diagnostics.

AN757

Extraction of Vitamin D Metabolites from Human Serum Using ISOLUTE® SLE+ in 96-Well Plate Format Prior to LC-MS-MS Analysis



Structure of 25-OH vitamin D₃.

Analytes

25-OH vitamin D₂ and 25-OH vitamin D₃.

Format

ISOLUTE® SLE+ 400 µL Supported Liquid Extraction plate, part number 820-0400-P01.

Matrices

Human Serum.

Sample Preparation Method

Dilute human serum (150 µL) with HPLC grade water:isopropanol (v/v, 50:50), (150 µL), add internal standard. Mix, cap and shake for 60 seconds.

Load the pre-treated serum (300 µL in total) onto the plate and apply a pulse of vacuum or positive pressure to initiate flow (3–5 sec). Allow the sample to absorb for 5 minutes.

Elute analytes with heptane (2 x 750 µL).

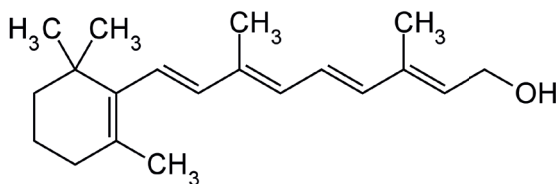
Post Extraction

Evaporate the eluate to dryness without heat and reconstitute in 100 µL of (v/v, 30:70) 2 mM ammonium formate (aq) with 0.1% formic acid: 2 mM ammonium formate (99% MeOH, 1% aq) with 0.1% formic acid before analysis.

Summary of Results

Recoveries were consistently greater than 90% with RSDs <10% and well within the standard deviations provided by DEQAS.





Notes

Notes

Notes

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